

SUBSTITUTE FORM PTO-1390

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER
13384-002001**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**U.S. APPLICATION NO. (If Known, see 37 CFR
1.5)

09/889874

INTERNATIONAL APPLICATION NO.
PCT/GB00/00219INTERNATIONAL FILING DATE
24 January 2000PRIORITY DATE CLAIMED
22 January 1999TITLE OF INVENTION
BIOLOGICAL CONTROL OF NEMATODES

APPLICANT(S) FOR DO/EO/US

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Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to promptly begin national examination procedures (35 U.S.C. 371(f)).
4. ☒ The US has been elected by the expiration of 19 months from the priority date (PCT Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)) (unsigned).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 16 below concern other documents or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A FIRST preliminary amendment.
☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☐ Other items or information:

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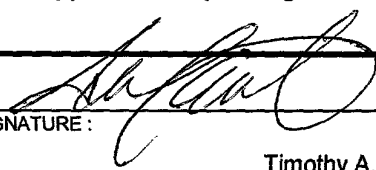
7-23-2001

Signature

Samantha Bell

Typed Name of
Person Signing

Samantha Bell

U.S. APPLICATION NO. (IF KNOWN) 09/889874		INTERNATIONAL APPLICATION NO. PCT/GB00/00219		ATTORNEY'S DOCKET NUMBER 13384-002001			
17. <input checked="" type="checkbox"/> The following fees are submitted:				CALCULATIONS PTO USE ONLY			
Basic National Fee (37 CFR 1.492(a)(1)-(5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1000 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710 International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) \$100 <div style="text-align: right;">ENTER APPROPRIATE BASIC FEE AMOUNT =</div>							
						\$860.00	
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Surcharge of \$130 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$0.00			
Claims	Number Filed	Number Extra	Rate				
Total Claims	34 - 20 = 4	4	x \$18	\$72.00			
Independent Claims	3 - 3 = 0	0	x \$80	\$0.00			
MULTIPLE DEPENDENT CLAIMS(S) (if applicable)			+ \$270	\$270.00			
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<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				\$0.00			
SUBTOTAL =				\$1,202.00			
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Timothy A. French FISH & RICHARDSON P.C. 225 Franklin Street Boston, MA 02110-2804 (617) 542-5070 phone (617) 542-8906 facsimile			<div style="text-align: center;">  SIGNATURE: </div> <div style="text-align: center;"> NAME: Timothy A. French </div> <div style="text-align: center;"> REGISTRATION NUMBER: 30,175 </div>				

14 / PRTS

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BIOLOGICAL CONTROL OF NEMATODES

TECHNICAL FIELD

The present invention relates to methods and materials for controlling nematodes.

PRIOR ART

Several thousand species of nematodes, sometimes called eel worms, are known. Numerous nematodes attack and parasitize humans and animals and cause disease. Additionally, several hundred species are known to feed on living plants. Certain of these are reviewed by Agrios in "Plant Pathology - 3rd Ed" Pub Academic Press Inc, see Chapter 15 therein.

Methods of controlling nematodes and their associated diseases include cultural practices; biological methods, e.g. use of resistant varieties; physical methods, e.g. heat; and use of chemical agents.

Patent application WO 92/19739 (Mycogen) relates to genes and gene fragments from *Bacillus thuringiensis* which have nematocidal activity. These generally encode crystal toxins from particular strains.

Patent application EP 0 303 426 (Mycogen) also relates to strains of *B. thuringiensis* which have nematocidal activity.

Patent application EP 0 171 381 (Monsanto) relates to particular soil bacteria which are capable of proliferating in an environment which is infested with

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nematodes such as pseudomonads which colonise the surface of plant roots. The basis for the controlling activity appears to stem from glycosidase enzymes which are hypothesised to directly inhibit the nematodes.

Notwithstanding these disclosures, there is an ongoing requirement for materials which have nematocidal activity, for instance for use in crop protection or nematode-mediated disease control.

Patent application PCT/WO 99/22598 (University of Reading) published 14 May 1999 claims a biopesticide for the control of insect pests or plant parasitic nematodes or both, which comprises as an effective agent a species of bacteria which is a symbiont of an entomopathogenic nematode.

DISCLOSURE OF THE INVENTION

The present inventors have established that species of bacteria which in nature are associated symbiotically with entomopathogenic nematodes, can in fact be utilised to control nematodes, and in preferred forms of the invention, to kill them. The bacteria themselves can be employed, or nematode control agents can be used which are derived from such bacteria. In one aspect of the invention, the present invention employs bacteria which are engineered and thus not naturally occurring, or nematode control agents which are derived from natural or non-natural bacteria.

It has been reported that certain bacterial species such as *Xenorhabdus* and *Photorhabdus* can be used to control insects, see e.g. PCT/WO 98/08388 of MAFF, PCT/WO 97/17432 of WARF, and PCT/WO 99/42589 of Novartis. An effect against nematodes had not previously been demonstrated.

The symbiotic bacteria used in the present invention are isolatable from nematodes or the insects which the nematodes attack, and differ fundamentally in terms of life-style and activity from those soil bacteria such as *B. thuringiensis* or pseudomonads which have previously been suggested as being nematocidal.

Indeed, *prima facie*, it seems highly unlikely that nematode symbiotes might possess nematocidal activity. However, in the light of the present disclosure, a number of possible explanations for the observed activity can be tentatively proposed. Firstly, in order to protect a nutrient supply from a dead insect, the bacteria might produce anti-nematocides to prevent saprophytic nematodes gaining access. Alternatively, to become a symbiont, the bacterial strains may have once been pathogens of these nematodes and evolved towards a less hostile symbiotic relationship. The nematocidal activity may be an evolutionary throwback from the original pathogenic relationship, in which case it may be expected to be widely present amongst bacteria which have evolved in this way.

A first aspect of the present invention is the use of bacterial strains to control a target nematode, characterised in that in nature the bacterial strain is associated symbiotically with an entomopathogenic nematode.

As discussed in more detail below, the bacterial strains may be used in the methods of the present invention *per se*, or they may be used as a source of nematode control agent. The nematode control agent can be derived directly, or be prepared and utilised through recombinant DNA techniques, optionally via a host cell.

The target nematode will generally be different to the nematode with which the bacterial strain is found symbiotically in nature.

By means of the present invention employing bacteria or a nematode control agent, it becomes possible to control nematodes, in the sense of, to prevent or retard the effect that the nematode has on other organisms such as animals or more preferably plants, or to reduce the number of nematodes or nematode eggs in an area of interest, or to alleviate or cure a disease caused by nematodes. Control may be at the level of larval nematodes or nematode eggs, or may inhibit the motion, feeding or infectivity of adult nematodes. Nematocidal control may be employed to kill the nematode target. Such controlling activity can be assessed as shown in the Examples below.

PREFERRED EMBODIMENTS

The present invention provides a composition for the control of parasitic nematodes which comprises as an effective agent a species of bacteria which is a symbiont of an entomopathogenic nematode, or engineered bacteria having such activity, or a nematode control agent derived from natural or engineered bacteria.

Correspondingly, the present invention also provides a method of nematode control employing such a composition.

The bacterial species is typically of the genera *Xenorhabdus* or *Photorhabdus*, preferably the genus *Xenorhabdus*, for instance the species *Xenorhabdus bovienii*. Examples of particularly preferred bacteria include:

Xenorhabdus bovienii strain H31 deposited with NCIMB under accession number NCIMB 40985 on 20 January 1999;

Xenorhabdus bovienii strain I73 deposited with NCIMB under accession number NCIMB 40986 on 05 November 1998; and

Xenorhabdus strain C42 deposited with NCIMB under accession number NCIMB 41004 on 05 November 1998.

The nematode control agent can be a peptide derived from a symbiont of an entomopathogenic nematode or an engineered bacterium has functional activity against a nematode. The peptide nematode control agent can be produced from a nucleic acid derived from a symbiont of an entomopath nematode or an engineered bacterium and which encodes such a peptide. The peptide can be an oligopeptide or a polypeptide, notably a protein. In one version, the nematode control agent is a toxin with toxic activity against nematodes, but the nematode control agent can have other activity.

The nucleic acids of this invention can be employed in a method of producing a peptide comprising the step of causing or allowing the expression from a nucleic acid of this invention in a suitable host cell.

The nucleic acid can comprise a natural nucleotide sequence or a degeneratively equivalent sequence, and functional variants thereof. Variants include homologous variants encoding a peptide which is a nematode control agent, the nucleic acid having 70% or more DNA sequence identity and/or the peptide having 70% or more amino acid sequence identity. Especially preferred nucleic acids in p 13-1f and p 14-2f and variants thereof.

The present invention extends to nucleic acids having a sequence which is a derivative by way of addition, insertion, deletion or substitution of one or more nucleotides. The nucleic acid can contain longer expressed sequences such that the nematode control agent is expressed as a fusion protein.

Nucleic acids complementary to the nucleic acid encoding a nematode

[illegible]

A method provided by this invention comprises the steps of:

- The hybridisation conditions can be selected to allow the identification of sequences having 70% or more sequence identity with the probe.

In one embodiment, the method comprises use of two primers to amplify a nucleic acid encoding a nematode control agent, at least one of the primers having a conserved nucleotide sequence of at least 15 nucleotides.

A method is further made possible by this invention comprising the steps of:

- (a) providing a preparation of nucleic acid from a bacterium,
- (b) providing a pair of nucleic acid molecule primers, at least one of which is a primer,
- (c) contacting nucleic acid in said preparation with said primers under

conditions for performance of PCR,

- (d) performing PCR and determining the presence or absence of an amplified PCR product.

Additionally, the invention provides a recombinant vector comprising a nucleic acid of this invention. The vector is preferably capable of replicating in a suitable host such as *E. coli* or in *Xenorhabdus*. The vector can be a baculovirus. In a preferred feature, the nucleic acid is operably linked to a promoter or other regulatory element for transcription in a host cell.

Vectors can further comprise any one or more of the following: a terminator sequence; a polyadenylation sequence; an enhancer sequence; a marker gene; a sequence encoding pesticidal material derived from *Bacillus thuringiensis*.

The vector can be a plant vector.

The vector of this invention can be introduced into a cell. Thus, a method for transforming a plant cell comprises the step of causing or allowing recombination between the vector and the plant cell genome to introduce the nucleic acid into the genome. The nucleic acid can be incorporated into chloroplast DNA, or into mitochondrial DNA.

Host cells comprising a vector are also part of this invention. The host cell can be a plant cell, which may be in a plant.

To this end, a method for producing a transgenic plant comprises the step of regenerating a plant from the transformed cell. In turn, plants of this invention extend to the progeny of such plants.

Examples of plants of this invention include crop species which can be

thuringiensis or pesticidal materials derived therefrom.

In a further aspect, there is provided an antibody or fragment thereof, or a polypeptide comprising the antigen-binding domain of the antibody, capable of specifically binding a peptide of this invention.

Such an antibody or fragment can be obtained by immunising a mammal with the peptide, and is useful in a method of identifying and/or isolating a nematode control agent comprising the step of screening candidate polypeptides with a polypeptide comprising the antigen-binding domain of the antibody of claim.

Some further aspects of preferred embodiments of the invention will now be discussed.

Bacterial strains

These can be derived from any entomopathogenic nematode. Preferred species are *Xenorhabdus* and *Photorhabdus*.

Potential sources of bacteria for use in the methods of the present invention may be identified by any preferred method. For instance, entomopathogenic nematodes can be isolated using an insect baiting technique such as that described by Bedding & Akhurst (1975) *Nematologia* 21: 215-227. Bacteria from nematodes identified as being pathogenic to the insect are isolated, cultured, and used as a source of nematocidal agent, e.g. by analogy with the methods used in the Examples below. Preferably *Xenorhabdus* or *Photorhabdus* species are used.

The preferred bacterial strains include ones which have the characteristics of

strain C42, I73 or H31 isolated by the present inventors. This *Xenorhabdus* strain has the following characteristics: rod shaped; motile; non-bio luminescent; blue on NBTA; produces antibiotics; resistant to ampicillin; forms circular colonies; has convex morphology; white colour.

This strain was presumptively identified as belonging to the genera *Xenorhabdus* since it was isolated from an insect killed by an entomopathogenic nematode and had the above characteristics. The strain has been deposited at the NCIMB (23 St Machar Drive, Aberdeen, AB24 3RY, Scotland) by the applicants under accession number NCIMB 41004 on 20 January 1999.

Further preferred strains of the present invention are two strains of *X. bovienii* designated H31 and I73 which have also been deposited under the terms of the Budapest Treaty at the NCIMB under the accession numbers NCIMB 40985 and 40986 respectively. These share characteristics of C42 in that they are rod-shaped; motile; non-bioluminescent; blue on NBTA; produce antibiotics; resistant to ampicillin; form circular colonies; and have convex morphology. The strains were identified as belonging to the species *X. bovienii* when compared to the *X. bovienii* type strain T228 using Restriction Analysis of the complete 16S rRNA gene and partial sequence analysis.

Target nematodes and diseases

The group of diseases described generally as helminthiasis is due to infection of an human or other animal host with parasitic worms known as helminths. Helminthiasis is a prevalent and serious economic problem in domesticated animals such as swine, sheep, horses, cattle, goats, dogs, cats and poultry. Among the helminths, the group of worms described as nematodes causes

widespread and often at times serious infection in various species of animals. The most common genera of nematodes infecting the animals referred to above are *Haemonchus*, *Trichostrongylus*, *Ostertagia*, *Nematodirus*, *Cooperia*, *Ascaris*, *Bunostomum*, *Oesophagostomum*, *Chabertia*, *Trichuris*, *Strongylus*, *Trichonema*, *Dictyocaulus*, *Capillaria*, *Heterakis*, *Toxocara*, *Ascaridia*, *Oxyuris*, *Ancylostoma*, *Uncinaria*, *Toxascaris*, *Caenorhabditis* and *Parascaris*. Certain of these, such as *Nematodirus*, *Cooperia*, and *Oesophagostomum*, attack primarily the intestinal tract, while others, such as *Dictyocaulus* are found in the lungs. Still other parasites may be located in other tissues and organs of the body.

The bacteria and encoded toxins of the invention may be used as nematocides for the control of the nematodes and diseases discussed above. More preferably, however, they are used to control soil and plant parasitic nematodes. Particular crop species which can be protected include tomatoes, potatoes, sugar beet, barley, soybean, peanut, onion, rye, wheat, corn, banana, raspberry, beans. Decorative and other plants may also be treated e.g. rose.

Target nematodes may be selected from the genera *Aphelenochoides*, *Anguina*, *Bursaphelenchus*, *Criconemella*, *Meloidogyne*, *Ditylenchus*, *Globodera*, *Helicotylenchus*, *Heterodera*, *Pratylenchus*, *Radopholus*, *Rotelynchus*, *Tylenchus*, *Trichodorus*, *Xiphenema*. A further organism used in certain of the Examples below is *Caenorhabditis elegans*. Other target organisms and plants are discussed by Agrios in "Plant Pathology - 3rd Ed" Pub Academic Press Inc, see Chapter 15 therein.

As stated above, the target nematode will generally be different to that with which the bacterial strain is found in nature.

Methods of use of bacteria

The bacteria may be used in any appropriate method which brings them into contact with the target nematode, preferably such that they, or their products, are ingested or absorbed by the target nematode.

In particular, regarding plants, the bacteria may be formulated in a variety of ways so as to enhance stability. For instance they may be employed in admixture with substrates to protect the cells.

The mixture can be spread over, ploughed into or otherwise mixed with nematode infected or potentially infected soil.

Regarding animals, bacteria intended for enteric inoculation can be mixed with carrier material that is suitable for ingestion by the intended animals.

Isolation of agent

Nematode control agents of the present invention, which may be proteinaceous, or nucleic acids encoding them, may be isolated and/or purified from the C42, I73 or H31 bacteria described above, in substantially pure or homogeneous form, or free or substantially free of other materials from the bacterial strain of origin. Where used herein, the term "isolated" encompasses all of these possibilities.

Methods of purifying proteins from heterogenous mixtures are well known in the art, e.g. selective precipitation, proteolysis, ultrafiltration with known molecular weight cut-off filters, ion-exchange chromatography, gel filtration, etc. A particularly useful initial technique in this regard is ultracentrifugation. Further methods which are known to be suitable for

protein purification are disclosed in "Methods in Enzymology Vol 182 - Guide to Protein Purification" Ed. M P Deutscher, Pub. Academic Press Inc. Other references which outline techniques commonly used by those of ordinary skill in the art include "Protein Purification - principles and practice" Pub. Springer-Verlag, New York Inc (1982), and by Harris & Angal (1989) "Protein purification methods - a practical approach " Pub. O.U.P. UK.

Nematocidal activity may be assessed using a spread assay as discussed below.

The C42, I73 or H31 agent may be wholly or partially synthetic. In particular they may be recombinantly produced from nucleic acid sequences which are not found together in nature (do not run contiguously) but which have been ligated or otherwise combined artificially.

For instance, in the Examples below, nucleic acid encoding toxin(s) from I73 has been expressed in hosts cells using a vector system. Amino acid sequences of 38 different putative I73 toxin(s) are set out in sequence Annex 1. These sequences are based on the nucleic acid sequence set out in Fig 2 ('chrim5'), a cosmid clone derived from I73 genomic DNA which conferred nematocidal activity upon *E. coli* cells into which it was introduced (i.e. significantly reduced nematode larval growth and development, and feeding). As detailed below, the entire amino acid sequence as set out in each case may not be required for nematocidal activity. In particular the portion up to the first Met in each sequence may be omitted, as may other portions which may not contribute to the nematocidal activity. Thus, not all the proteins or genes may be required for nematocidal activity, and usually there will be one or more principal proteins, though others may play supporting roles such as in enhancing the activity or encoding other nematocidal activities.

Thus isolated nematocidal agents comprising a polypeptide containing all, or a nematocidal fragment, of any of the depicted I73 sequences, form one aspect of the present invention. Preferred agents include those encoded by p14-2f and p13-1f. Other active variants of these sequences are also encompassed as described below.

Candidate agents for use in this invention to control nematodes extend to those from the bacteria described in PCT/WO 99/22598, as well as the insecticidal toxins and bacteria of PCT/WO 99/42589, PCT/WO 98/08388 and PCT/WO 97/17432, the disclosures of which are incorporated by reference.

Nucleic acids and variants

In one aspect of the present invention there is provided a nucleic acid molecule encoding a nematode control agent of the present invention, for example a toxin, as described above.

The nucleic acid may be derived from the sequence shown in Fig 2 or the complement (or degenerate equivalent) thereof. This sequence (cHRIM5) was itself derived from I73 and identified by its unexpected nematocidal activity. Regions of this sequence believed to correspond to genes of the present invention are described in Fig 3. Isolated nucleic acids comprising one or more of these regions which encode a nematocidal activity are particularly preferred.

In the light of the present disclosure, further nucleic acids of the present invention may be isolated using PCR or southern blotting or other techniques well known to those skilled in the art. This requires the use of two primers to specifically amplify target nucleic acid, so preferably two

nucleic acid molecules with sequences characteristic of the C42, H31 or most preferably an I73 toxin isolated as above are employed. Using RACE PCR, only one such primer may be needed (see "PCR protocols: A Guide to Methods and Applications", Eds. Innis et al, Academic Press, New York, (1990)).

Thus a method involving use of PCR in obtaining nucleic acid according to the present invention may include:

- (a) providing a preparation of bacterial nucleic acid,
- (b) providing a pair of nucleic acid molecule primers suitable for PCR, at least one of said primers being a primer based on a toxin from C42, H31 or I73,
- (c) contacting nucleic acid in said preparation with said primers under conditions for performance of PCR,
- (d) performing PCR and determining the presence or absence of an amplified PCR product. The presence of an amplified PCR product may indicate identification of a variant.

In a further aspect of the present invention there are disclosed nucleic acids which are variants of the C42, I73 or H31 toxin. A variant nucleic acid molecule shares homology (or identity) with all or part of the C42, H31, or most preferably I73 sequence discussed above.

Preferably sequence comparisons are made using FASTA and FASTP (see Pearson & Lipman, 1988. Methods in Enzymology 183: 63-98). Parameters are set, using the default matrix blosum62, as follows:

Gapopen (penalty for the first residue in a gap): -12 for proteins / -16 for DNA

Gapext (penalty for additional residues in a gap): -2 for proteins / -4 for DNA

KTUP word length: 2 for proteins / 6 for DNA.

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Homology (similarity or identity) may be at the nucleotide sequence and/or encoded amino acid sequence level. Preferably, the nucleic acid and/or amino acid sequence shares at least about 70%, 75%, 80%, or 85% homology, most preferably at least about 90%, 95%, 96%, 97%, 98% or 99% homology.

Another method for assessing homology at the nucleic acid level is by hybridization screening. One common formula for calculating the stringency conditions required to achieve hybridisation between nucleic acid molecules of a specified sequence homology is shown in Molecular Cloning: a Laboratory Manual: 2nd edition, Sambrook et al, 1989, Cold Spring Harbor Laboratory Press:

$$T_m = 81.5^{\circ}\text{C} + 16.6\text{Log} [\text{Na}^+] + 0.41 (\% \text{ G+C}) - 0.63 (\% \text{ formamide}) - 600/\text{\#bp}$$
in duplex

As an illustration of the above formula, using $[\text{Na}^+] = [0.368]$ and 50-% formamide, with GC content of 42% and an average probe size of 200 bases, the T_m is 57°C . The T_m of a DNA duplex decreases by 1 - 1.5°C with every 1% decrease in homology. Thus, targets with greater than about 75% sequence identity would be observed using a hybridization temperature of 42°C . Such a sequence would be considered substantially homologous to the nucleic acid sequence of the present invention.

Variants of the present invention can be artificial nucleic acids.

Alternatively they may be novel, naturally occurring, nucleic acids, isolatable using the information disclosed herein. Thus a variant may be a distinctive part or fragment (however produced) corresponding to a portion of the C42, I73 or H31 toxin. The fragments may encode particular functional parts of the agent or they may be used for probing for, or amplifying, sequences corresponding to C42, I73 or H31 toxin. Sequence variants which occur naturally may include homologs of the C42, I73 or H31 toxin from other

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bacteria, including nematode-symbionts. Artificial variants (derivatives) may be prepared by those skilled in the art, for instance by site directed or random mutagenesis (i.e. nucleotide addition, deletion or substitution, optionally to lead to amino acid addition, deletion or substitution) or by direct synthesis. Preferably the variant nucleic acid is generated either directly or indirectly from an original nucleic acid encoding the C42, I73 or H31 toxin.

Changes may be desirable for a number of reasons, including introducing or removing the following features. Sites which are required for pre- or post-translation modification. Changes for codon usage preferences to enhance gene expression in different organisms. Leader or other targeting sequences (e.g. membrane or golgi locating sequences) may be added to the expressed protein to determine its location following expression. All of these may assist in efficiently cloning and expressing an active polypeptide in recombinant form. Other desirable mutation may be random or site directed mutagenesis in order to alter the activity (e.g. host specificity) or stability of the encoded polypeptide. Changes may be by way of conservative variation, i.e. substitution of one hydrophobic residue such as isoleucine, valine, leucine or methionine for another, or the substitution of one polar residue for another, such as arginine for lysine, glutamic for aspartic acid, or glutamine for asparagine. Also included are active (nematocidal) variants having non-conservative substitutions.

Variant nucleic acids encompass all of these possibilities. When used in the context of polypeptides or proteins they indicate the encoded expression product of the variant nucleic acid i.e. variants of C42, I73 or H31 toxin e.g. variants of the I73 toxin sequences disclosed hereinafter.

Vectors and production of host cells

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In one aspect of the present invention, the nucleic acid encoding the nematode control agent is provided in the form of a recombinant and preferably replicable vector.

Generally speaking, those skilled in the art are well able to construct vectors and design protocols for recombinant gene expression. Suitable vectors can be chosen or constructed, containing appropriate regulatory sequences, including promoter sequences, terminator fragments, polyadenylation sequences, enhancer sequences, marker genes and other sequences as appropriate. For further details see, for example, Sambrook et al (1989) *supra*.

The permitted vectors include, *inter alia*, any plasmid, cosmid, phage or *Agrobacterium* binary vector in double or single stranded linear or circular form which may or may not be self transmissible or mobilizable, and which can transform a prokaryotic or eukaryotic host either by integration into the cellular genome or exist extrachromosomally, e.g. an autonomous replicating plasmid with an origin of replication. Illustratively integration can occur into chloroplast DNA or into mitochondrial DNA.

Preferably the nucleic acid in the vector is under the control of, and operably linked to, an appropriate optionally inducible promoter or other regulatory elements for transcription in a host cell such as a microbial, e.g. bacterial, yeast, filamentous fungal or plant cell. The vector may be a bi-functional expression vector which functions in multiple hosts. In the case of genomic DNA, this may contain its own promoter or other regulatory elements and in the case of cDNA this may be under the control of an appropriate promoter or other regulatory elements for expression in the host cell. The vectors and host cells into which they are introduced may be used to clone or otherwise

fragments. Hosts can also be used to generate quantities of toxin which can be employed in situ in suitable treated cells, or alternatively with suitable hosts, e.g., *Pseudomonas* viable microbes can be applied to the sites of nematodes where they will proliferate and where they or their products can be ingested by the nematodes. Higher organisms, preferably plants, can also be engineered with the toxin. The result in each case is a control of the nematodes. A host may be selected that can tolerate harsh environmental conditions and then grow when they improve, as illustrated by *Bacillus* species where the spores can exist under environmental extremes.

Characteristics of interest for use as a nematocide microcapsule i.e. a vehicle for the active agent include protective qualities for the nematocide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to nematodes for ingestion; ease of killing and fixing without damage to the toxin; and the like.

Treated host cells

Where the cell is treated, the cell will usually be intact and be substantially proliferative form when treated, rather than in a spore form, although in some instances spores may be employed. Treatment of the microbial cell, e.g. a microbe containing the bacterial toxin gene or gene fragment, can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the toxin, nor diminish the cellular capability in protecting the toxin.

Viable hosts

Where the toxin gene or gene fragment is introduced via a suitable vector into a microbial host, and said host is applied to the environment in a living state, it is preferable that microorganism hosts are selected which are known to occupy the phytosphere (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment (crop and other insect habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the nematocide from environmental degradation and inactivation.

A large number of microorganisms are known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots) of a wide variety of important crops. These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., genera *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Xanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylophilus*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*, *Arthrobacter*, *Azotobacter*, *Leuconosroc*, and *Alcaligenes*; fungi, particularly yeast, e.g., genera *Saccharomyces*, *Cryptococcus*, *Kluyveromyces*, *Sporobolomyces*, *Rhodororula*, and *Aureobasidium*.

Plants as hosts

Nucleic acid encoding the nematocides of the present invention can be introduced into plant cells using any suitable technology, such as a disarmed Ti-plasmid vector carried by *Agrobacterium* exploiting its natural gene transfer ability (EP-A-270355, EP-A-0116718, NAR 12(22) 8711 - 87215 1984), particle or microprojectile bombardment (US 5100792, EP-A-444882,

EP-A-434616) microinjection (WO 92/09696, WO 94/00583, EP 331083, EP 175966, Green et al. (1987) Plant Tissue and Cell Culture, Academic Press), electroporation (EP 290395, WO 8706614 Gelvin Debeyser) other forms of direct DNA uptake (DE 4005152, WO 9012096, US 4684611), liposome mediated DNA uptake (e.g. Freeman et al. Plant Cell Physiol. 29: 1353 (1984)), or the vortexing method (e.g. Kindle, PNAS U.S.A. 87: 1228 (1990d). Physical methods for the transformation of plant cells are reviewed in Oard, 1991, Biotech. Adv. 9: 1-11.

Agrobacterium transformation is widely used by those skilled in the art to transform dicotyledonous species. It has also been used with filamentous fungi (see de Groot et al, 1998, Nature Biotechnology 16: 839-842).

Recently, there has also been substantial progress towards the routine production of stable, fertile transgenic plants in almost all economically relevant monocot plants (see e.g. Hiei et al. (1994) The Plant Journal 6, 271-282)). Microprojectile bombardment, electroporation and direct DNA uptake are preferred where *Agrobacterium* alone is inefficient or ineffective. Alternatively, a combination of different techniques may be employed to enhance the efficiency of the transformation process, e.g. bombardment with *Agrobacterium* coated microparticles (EP-A-486234) or microprojectile bombardment to induce wounding followed by co-cultivation with *Agrobacterium* (EP-A-486233).

Generally speaking, following transformation, a plant may be regenerated, e.g. from single cells, callus tissue or leaf discs, as is standard in the art. Almost any plant can be entirely regenerated from cells, tissues and organs of the plant. Available techniques are reviewed in Vasil et al., Cell Culture and Somatic Cell Genetics of Plants, Vol I, II and III, Laboratory Procedures and Their Applications, Academic Press, 1984, and Weissbach and

Weissbach, Methods for Plant Molecular Biology, Academic Press, 1989.

The generation of fertile transgenic plants has been achieved in the cereals rice, maize, wheat, oat, and barley (reviewed in Shimamoto, K. (1994) Current Opinion in Biotechnology 5, 158-162.; Vasil, et al. (1992) Bio/Technology 10, 667-674; Vain et al., 1995, Biotechnology Advances 13 (4): 653-671; Vasil, 1996, Nature Biotechnology 14 page 702).

Combination nematocides

In further embodiments of the invention, bacteria associated with entomopathogenic nematodes or the toxins or products discussed above are used in conjunction with other nematocidal bacteria such as *B. thuringiensis* strains (e.g. from WO 92/19739) or pesticidal materials derived therefrom.

Materials for use in the present invention

The present invention also embraces materials for use in the methods above. These materials include the novel bacterial strains which are associated symbiotically with an entomopathogenic nematode and which are capable of controlling a target nematode. In particular the invention encompasses strain C42, I73 or H31 in isolated or substantially isolated form, or strains having the characteristics of C42, I73 or H31 (including nematocidal activity assessed as below).

Also embraced are compositions and formulations of these bacteria. These may comprise or consist of wettable powders, granules or dusts, mixed with various inert materials, such as inorganic minerals (phyllosilicates, carbonates, sulfates, phosphates, methylcellulose, xanthan gum and the like) or botanical materials (powdered corncobs, rice hulls, walnut shells,

peat moss, vermiculite, soil, seeds, other plant tissue and the like). The formulations may include spreader-sticker adjutants, stabilizing agents or surfactants. Liquid formulations may be aqueous-based or non-aqueous and employed as foams, gels, suspensions, emulsifiable concentrates, or the like. The ingredients may include rheological agents, surfactants, emulsifiers, dispersants, or polymers.

Bacteria may be mixed with other material while in freeze-dried form, encapsulated in biodegradable or water-soluble material, or otherwise treated to prolong their viability or decrease their levels of metabolic activity during handling. If desired, the carrier material may contain assimilatable nutrient sources to support proliferation of the bacteria.

Also included are purified or substantially purified nematocidal agents (particularly proteinaceous agents) isolated or isolatable from the strains or host cells discussed above.

Thus the invention further discloses nematocidal compositions comprising one or more agents as described above. Such compositions preferably further comprise other nematocidal materials from other *Xenorhabdus* species or non-*Xenorhabdus* species. These other materials may be chosen such as to have complementary properties to the agents described above, or act synergistically with it.

Toxins of the invention for use with animals can be adapted to be administered orally in a unit dosage form such as a capsule, bolus or tablet, or as a liquid drench when used as an anthelmintic in mammals, and in the soil to control plant nematodes. The drench is normally a solution, suspension or dispersion of the active ingredient, usually in water, together with a suspending agent such as bentonite and a wetting agent or like

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excipient. Generally, the drenches also contain an antifoaming agent. Drench formulations generally contain from about 0.001 to 0.5% by weight of the active compound. Preferred drench formulations may contain from 0.01 to 0.1% by weight, the capsules and boluses comprise the active ingredient admixed with a carrier vehicle such as starch, talc, magnesium stearate, or dicalcium phosphate. Where it is desired to administer the toxin compounds in a dry, solid unit dosage form, capsules, boluses or tablets containing the desired amount of active compound usually are employed. These dosage forms are prepared by intimately and uniformly mixing the active ingredient with suitable finely divided diluents, fillers, disintegrating agents and/or binders such as starch, lactose, talc, magnesium stearate, vegetable gums and the like. Such unit dosage formulations may be varied widely with respect to their total weight and content of the antiparasitic agent, depending upon the factors such as the type of host animal to be treated, the severity and type of infection and the weight of the host.

When the active compound is to be administered via an animal feedstuff, it is intimately dispersed in the feed or used as a top dressing or in the form of pellets which may then be added to the finished feed or, optionally, fed separately. Preferably, a carrier for feed administration is one that is, or may be, an ingredient of the animal ration. Suitable compositions include feed premixes or supplements in which the active ingredient is present in relatively large amounts and which are suitable for direct feeding to the animal or for addition to the feed either directly or after an intermediate dilution or blending step. Typical carriers or diluents suitable for such compositions include, for example, distillers' dried grains, corn meal, citrus meal, fermentation residues, ground oyster shells, wheat shorts, molasses solubles, corn cob meal, edible bean mill feed, soya grits, crushed limestone and the like.

Alternatively, the antiparasitic compounds may be administered to animals parenterally, for example, by intraluminal, intramuscular, intratracheal, or subcutaneous injection, in which event the active ingredient is dissolved or dispersed in a liquid carrier vehicle. For parenteral administration, the active material is suitably admixed with an acceptable vehicle, preferably of the vegetable oil variety, such as peanut oil, cotton seed oil and the like. Other parenteral vehicles, such as organic preparations using solketal, glycerol, formal and aqueous parenteral formulations, are also used. The active compound or compounds are dissolved or suspended in the parenteral formulation for administration; such formulations generally contain from 0.005 to 5% by weight of the active compound.

Further aspects of the invention include nucleic acids, vectors and host cells containing a heterologous construct according to the present invention, especially a plant or a microbial cell.

Such microbial cells may be treated as described in the methods above. Examples of chemical reagents are halogenating agents. Other suitable techniques include treatment with aldehydes, such as formaldehyde and glutaraldehyde; anti-infectives, such as zephiran chloride and cetylpyridinium chloride; alcohols, such as isopropyl and ethanol; various histologic fixatives, such as Bouin's fixative and Helly's fixative (See: Humason, Gretchen L., Animal Tissue Techniques, W.H. Freeman and Company, 1967); or a combination of physical (heat) and chemical agents that preserve and prolong the activity of the toxin produced in the cell when the cell is administered to the host animal. The method of inactivation or killing retains at least a substantial portion of the bio-availability or bioactivity of the nematode control agent.

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In all of the compositions discussed above, the nematocide concentration may vary widely depending upon the nature of the particular formulation, particularly whether it is a concentrate or to be used directly. The nematocide will be present in at least 1% by weight and may be 100% by weight. The dry formulations will have from about 1-95% by weight of the nematocide while the liquid formulations will generally be from about 16% by weight of the solids in the liquid phase. The formulations will generally have from about 10^2 to about 10^{10} cells/mg, more preferably 10^7 to about 10^9 cells/mg. These formulations will be administered at about 50 mg (liquid or dry) to 1 kg or more per hectare. The formulations can be applied to the environment of the nematodes, e.g., plants, soil or water, by spraying, dusting, sprinkling, or the like.

In addition to the above the invention includes plant cells which have been transformed with the genes of the present invention, and plants which include such plant cells.

EXAMPLES OF THE INVENTION

The invention will now be further described with reference to the following non-limiting Figures and Examples. Other embodiments of the invention will occur to those skilled in the art in the light of these.

FIGURES

Fig 1 shows the cHRIM5 cosmid vector and subclones used for sequencing, as described in Example 6.

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Fig 2 shows the sequence of cHRIM5 (1-37544 bps).

Fig 3 shows the position and orientation of ORFs in the cHRIM5 sequence.

Fig 4 shows deletions of cHRIM5 tested for nematocidal activity.

Fig 5 illustrates cloning of nematocidal activity in PLEX.

Example 1 - Source of strains C42, I73 and H31

Strain C42 was obtained using an insect entrapment method. Insects which were killed on the surface of a soil sample were observed under a microscope at high magnification. Any that contained high numbers of bacteria and not fungal hyphae were presumed to have been killed by insect parasitic nematodes. The identified presence of nematodes also aids this identification step, but it is not essential. These samples were plated on to NBTA media (see Poinar & Thomas, 1984 Nematodes p238-280 in "Laboratory guide to insect pathogens and parasites" Eds. Poiner & Thomas, Pub. Plenum Press, New York). Any colonies that developed that had characteristic features (e.g. morphology, size, colour) of *Xenorhabdus* or *Photorhabdus* strains were selected. Non-luminescent colonies were presumptively identified as *Xenorhabdus*. The identity of those having nematocidal activity as assessed in Example 3, is further confirmed using 16s rRNA sequence data (see Brunel et al 1997, Applied and Environmental Microbiology 63,2: 574-580).

I73 and H31 strains were obtained in a similar way to strain C42 but they were identified as belonging to the species *X. bovienii* when compared to the *X. bovienii* type strain T228 using Restriction Analysis of the complete 16S

rRNA gene (see Brunel et al, 1997 Applied and Environmental Microbiology: 574-580), and partial 16s ribosomal RNA sequence analysis.

Example 2 - Cell growth and preservation

Subcultures of the *Xenorhabdus* species C42, I73 and H31 were used to inoculate three 9 cm diameter petri dishes containing L agar (10g tryptone, 5 g Yeast Extract, 5 g NaCl and 15 g agar per lt). Plates were incubated for 48 hrs at 26°C and the resulting growth harvested by scraping off bacterial cells and thoroughly resuspending in 40 mls of 5% w/v lactose. The cells were washed once by centrifugation (5000 x g for 10 mins), resuspended in 10 mls of 5% w/v lactose, dispensed into 1 ml aliquots and freeze dried (-60°C for 48 hrs) for medium term storage at 2°C. Other stocks were re-suspended in nutrient broth containing 10% w/v glycerol (Protect) and frozen at -70°C.

Example 3- Activity of cells against *Caenorhabditis elegans*

The bioassays were performed by allowing *C. elegans* to feed on live bacterial cell suspensions spread over the surface of Luria broth agar (Luria broth containing 1.2%w/v agar) in segmented square petri dishes (2.0 x 2.0 cm per test well). A minimum of three test wells, each containing 50-100 nematodes were used for each test. Mortalities were recorded after 3 days at 18°C.

C. elegans was cultured on *Escherichia coli* at 18°C on 9 cm diameter LB agar plates. Once the nematodes had colonised the complete plate they were re-subbed on a fresh plate to maintain stocks and the remainder re-suspended in 40 ml LB. The tube was allowed to stand for 15 min and the nematodes settled to the bottom. The concentrated nematodes were removed using a

sterile pipette and placed in 40 mls of fresh LB. The process was repeated 5 more times to wash the nematodes away from the *E. coli* cells. The nematodes were then diluted so that approximately 50 nematodes were present in 50 μ l of LB.

The *Xenorhabdus* cells used were cultured in LB at 30°C/100 rpm for 24 hours and 50 μ l spread on to the surface of each test well. The control *E. coli* cells were treated in a similar way but incubated at 37°C for growth. After application the wells were air dried for 30 min, and 50 μ l of the nematode suspension placed in each well. Again the wells were air dried for 30 min. Plates were incubated at 18°C with 80% relative humidity for 3 days.

Xenorhabdus spp. C42, H31 and I73 gave 95% mortality, as compared with no significant effect for certain other *Xenorhabdus* bacterial strains and *E. coli*. Thus these results clearly show that cells from *Xenorhabdus* C42, H31 and I73 are an effective nematocide.

Example 4 - Cloning of nematode active gene from I73

Total DNA was isolated from I73 using a Quiagen genomic DNA purification kit (cat no. 10243). To isolate DNA, cells were grown in Luria broth (10g tryptone, 5g yeast and 5g NaCl per lt) at 26°C with shaking at 200 rpm to an optical density of 1.5 A600. Cells were harvested by centrifugation at 4000 x g and the DNA isolated using Quiagen 100/G tips, as per manufacturer's instructions. The purified DNA was stored at -20°C in TE buffer (10 mM Tris, 1 mM EDTA, pH 8.0).

To obtain a representative I73 library, total DNA was partially digested with *Sau*3A. Approximately 25 μ g of DNA was incubated at 37°C with 0.25 units

of enzyme. At intervals of 5, 15, 30, 45 and 60 minutes, samples were removed and heated at 65°C for 10 minutes. To determine the size of the resulting DNA fragments, the samples were separated on a 0.5% (w/v) agarose gel. The samples containing a dominant DNA fragment size of between 30 and 50 Kb were combined and treated with shrimp alkaline phosphatase (Boehringer) for 20 minutes at 37°C. The DNA was ligated into the *Bam*HI site of the Stratagene cosmid vector Supercos1 (scos) and packaged into the *Escherichia coli* strain XL Blue 1, using a Gigapack II packaging kit (Stratagene) following the manufacturer's instructions.

To identify individual cosmid clones with activity to *C. elegans*, single colonies were grown in individual wells of segmented square petri dishes on Luria agar, containing 50 µg/ml ampicillin at 30°C for 24 hours. To each well, approximately 50, mainly L4 and adult *C. elegans* larvae were added in 50 µl of Luria broth. The dishes were incubated at 18°C and examined after 6 days for nematode development.

A total of 600 clones were examined and one coded cHRIM5 was found, which caused significant reduction in larval numbers, with no live L4 and adult larvae observed compared to on average, greater than 40 in all other clones tested.

Example 5 - Activity of cHRIM5, C42, H31 and I73 against *C. elegans*

Clone cHRIM5 was grown in 50 mls LB containing 50 µg of ampicillin per ml at 30°C/200 rpm for 40 hours. C42, H31 and I73 were grown in 50 mls LB at 26°C for 48 hours/200 rpm. Cultures were centrifuged at 4000 x g for 10 minutes, washed once and resuspended in 5 mls of PBS (0.05 mM phosphate buffer, 0.125M NaCl). To determine activity, 300 µl of cells were added in triplicate, to 1.2 ml of PBS containing 25, mainly L4 and adult *C.*

elegans larvae in multi well square dishes. As a control, an equivalent amount of XL 1 Blue *E. coli* cells containing Supercos 1 were used to determine nematode survival. The assays were incubated at 18 °C for 7 days before approximate nematode counts and observations were made.

Activity of cells on *C. elegans*

Cell line	No. and size of larvae/square	Cell turbidity
XL 1 Blue/Supercos 1	>100 (all stages)	Clear
XL 1 Blue/cHRIM5	<20(mainly small, L1,2 &3)	Cloudy
C42	<10	Cloudy
H31	<10	Cloudy
I73	<10	Cloudy

Thus cHRIM5, C42, H31 and I73 all gave a reduction in nematode numbers, and in particular cHRIM5 cells significantly reduced larval growth and development. All four strains caused a reduction in feeding (as indicated by the cloudy cell suspensions).

Example 6 - DNA and protein sequences

Plasmid and cosmid DNA for cloning was prepared using the QIAGEN midi system (tip 100, cat. No 12143). Cells were grown in Luria broth (Merck) at 37°C with shaking at 200 rpm for 18 hours. Cells were harvested by centrifugation at 6,000 x g and the DNA isolated as per manufacturers instructions. Restriction digestion (Roche, Life Technologies), dephosphorylation (Roche) and ligation (Life Technologies) were carried out using manufacturer's recommended conditions and as outlined by Sambrook et al. Transformation was accomplished using electrocompetant cells and a

BIO-RAD Gene pulser set at 12.5V cm⁻². Two µl of DNA was used to electroporate 80 µl of early log phase *E. coli* DH5 alpha cells washed 3 times in sterile water (centrifugation at 6000 x g for 5 mins) and resuspended in 1/100th the original volume in 10% (v/v) glycerol. Luria agar containing either kanamycin or ampicillin at 50 µg ml⁻¹ were used to select clones where appropriate.

DNA sequence analysis of cHRIM5 was completed by sequencing a number of sub clones and primer walking, see figure 1 for the supercos vector, where the numbers are kbp. The sub clones used are as follows:

code	cHRIM5 treatment	vector used or remaining
A-380	<i>Hind</i> III digestion and self-ligation	deleted scos
B-387	<i>Bam</i> HI digestion and self-ligation	pUC 19- <i>Bam</i> HI digestion
C-381	<i>Sal</i> I- <i>Bam</i> HI digestion	scos
E-391	<i>Sal</i> I- <i>Bam</i> HI digestion	pUC 19- <i>Sal</i> I <i>Bam</i> HI digestion
F-392	<i>Sal</i> I- <i>Bam</i> HI digestion	pUC 19- <i>Sal</i> I <i>Bam</i> HI digestion

Sub clone A-380 was constructed by digesting cHRIM5 DNA with the restriction enzyme *Hind*III and re-ligating fragments, this clone contains a deletion of the insert and scos cosmid DNA as the vector. Sub clone B-387 is a *Bam*HI digestion of cHRIM5 cloned into the plasmid pUC19 also cut with *Bam*HI and dephosphorylated. Sub clone C-381 was obtained by digesting cHRIM5 DNA with *Bam*HI and re-ligating the fragments, this clone contains the scos cosmid as the vector. Clones E-391 and F-392 were obtained by cutting cHRIM5 DNA with *Sal*I and *Bam*HI and ligating these fragments into the vector pUC19 also cut with these enzymes.

Sequencing was conducted using the artificial transposon AT2 (supplied by Perkin-Elmer-Applied Biosystems, Primer Island Transposition kit, cat No.

403015) using the cosmid cHRIM5 and all sub-clones as target DNA. One μg of cHRIM5 DNA was incubated with the transposon AT2 for 1 hour at 30°C in a final volume of $20\ \mu\text{l}$. After incubation the reaction was stopped by adding $5\ \mu\text{l}$ of 0.25M EDTA, 1% (w/v) SDS, and heat treatment at 65°C for 30 mins. The DNA was desalted by dialysis against water. One μl of the reaction mix was used to electroporate $80\ \mu\text{l}$ of early log phase *E. coli* DH5 alpha cells. Colonies were selected on LB media containing $50\ \mu\text{g/ml}$ trimethoprim. Once inserted the transposon mutants were used to provide a range of positions of primer sites at random intervals throughout the clones. The two primers PI+ and PI- near the end of the transposon were used to generate sequence data. In addition standard primers for the pUC19 and scos vectors were used to generate sequence data at the ends of each clone. DNA for sequencing was prepared using the QIAGEN ion exchange media (qiawell8, cat. No. 17122). Clones were grown in $1\ \text{ml}$ of Luria broth containing trimethoprim ($50\ \mu\text{g ml}^{-1}$) for 18 hours. Cells were centrifuged at $13,000 \times g$ for 5 mins and resuspended in $350\ \mu\text{l}$ of buffer P1. After 5 mins $350\ \mu\text{l}$ of buffer P2 was added and the samples incubated for 5 mins at room temperature. To this $350\ \mu\text{l}$ of buffer P3 was added and the samples left on ice for 15 mins. After centrifugation at $13,000 \times g$ for 15 mins the samples were loaded on the Qiagen column under vacuum, and washed with buffer QC. DNA was eluted with buffer QF ($500\ \mu\text{l}$) at 50°C and isopropanol precipitated ($0.8\ \text{vol}$). After centrifugation at $13,000 \times g$ for 30 min, DNA was washed with 70% (v/v) ethanol and air dried for 10 mins. The final pellet was resuspended in $10\ \mu\text{l}$ of water. Cycle sequencing reactions using the Perkin-Elmer Applied Biosystems division Big Dye reaction kit (cat No. 4303149) were prepared using standard conditions for plasmid and cosmid sequencing. Samples were analysed on ABI Automated Sequencers. DNA sequences were assembled using the DNA* software. The complete sequence of cHRIM5 was obtained by primer walking to join the final DNA contigs together. The final sequence of cHRIM5ed2 is shown

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in Figure 2. Analysis of the DNA using the software Clone indicated a number of ORF illustrated in Figure 3 and 4. Corresponding protein sequences are also presented at Annex I.

Example 7 - Fragments that encode nematocidal activity

To identify smaller fragments that encoded nematocidal activity, a series of sub-cloning experiments were performed using *E. coli* DH5 alpha. Qiagen midi and miniprep methods, restriction and ligations were used as for previous examples. Nematicidal activity of all constructs was determined as described in Example 4. In Figure 4, we show the deletions of cHRIM5 tested for nematocidal activity. Restriction sites and genes are indicated. Size in base pairs indicated on the map line. A cHRIM5, B cHRIM6, C cHRIM7, D cHRIM8, E cHRIM8, F cHRIM10, G *NdeI* deletion of cHRIM8, H Approximate positions (arrows) of three AT2 transposon insertions (tn58, tn26, tn43) in cHRIM9.

The cosmid cHRIM5 (figure 4A) was digested with the enzyme *SaII* and religated. The resulting sub clone cHRIM6, illustrated in Figure 4B showed nematocidal activity. cHRIM6 was digested with the enzyme *SmaI* and religated, producing sub-clone cHRIM7 (Figure 4C). cHRIM7 was digested with *BglII* and the kanamycin resistance gene block (*nptII*, Pharmacia) cut with *BamHI* was ligated into it. After selection on LB containing kanamycin (50µg ml⁻¹) and ampicillin (50µg ml⁻¹) the clone was digested with *SaII* and religated, in effect creating a deletion from the *SaII* site to the *BglII* site of cHRIM6 to generate cHRIM8 (figure 4D). By cutting cHRIM8 with *NruI* a further deletion was made to create cHRIM10 (figure 4F). All the above clones maintained nematocidal activity.

Deletion of cHRIM8 with *NdeI*, removed a portion of the p14-2f gene (figure

4G), this reduced nematocidal activity. This indicates that the p14-2f gene or protein are important for nematocidal activity. Transposon mutagenesis of cHRIM9 (a clone very similar to cHRIM7 but deleted with *NarI* rather than *SmaI*) with the artificial transposon AT2 (Perkin Elmer Applied Biosystems) resulted in a number of inserts within this clone (figure 4H). Insert cHRIM9-tn43 was restriction mapped to an approximate position of bp 20,700 (on cHRIM5) within the p20-9r gene, this mutant retained nematocidal activity. This indicates that this gene is not essential for activity. Insert cHRIM9-tn58 mapped to an approximate position of bp 13,400 (on cHRIM5), within the p13-1f gene, nematocidal activity was reduced. This indicates that this gene, region of DNA or the blocking effect of the transposon in this position is important for activity. Insert cHRIM9-tn26 was restriction mapped to approximate position of bp 15,000 (on cHRIM5) within the p14-2f gene, nematocidal activity was reduced. This indicates that this gene, region of DNA or the blocking effect of the transposon in this position is important for activity.

Clone cHRIM6-tn43 was digested with *Bgl*III and *Nof*I and cloned into the vector PLEX (Invitrogen cat. No. K450-01) cut with *Bam*HI and *Nof*I. The *E. coli* strain used was GI742 supplied by Invitrogen. The resulting plasmid insert (PLEX-*Bgl*III/tn43, Figure 5) places the p14-2f and p13-1f genes under the control of the bacteriophage Lambda P_L promoter. Figure 5 illustrates the cloning of DNA encoding nematocidal activity in the expression vector PLEX, where: A, plasmid clone; B, insert and gene locations; Tpr, trimethoprim resistance; Apr, ampicillin resistance; P_L , bacteriophage lambda P_L promoter; *, plasmid joins to form a circular molecule; **, incomplete genes. Selection of colonies on RMG media (described in the Invitrogen manual) containing ampicillin ($50 \mu\text{g ml}^{-1}$) and trimethoprim ($50 \mu\text{g ml}^{-1}$) prevents expression from the P_L promoter. Colonies were then cultured on LB containing Trimethoprim ($50 \mu\text{g ml}^{-1}$) in 2.0 cm^2 wells for

nematocidal tests. The clone was active. This indicates that genes within this fragment have nematocidal activity. The clone PLEX-*Bgl*II/tn43 was digested with *Cla*I and religated, this resulted in a deletion of part of the p13-1f gene, this clone had reduced nematocidal activity indicating the importance of this gene.

All these results indicate that the genes and gene products of p13-1f and p14-2f are important for nematocidal activity. Other smaller genes within the *Bgl*II to *Nru*I sites of cHRIM10 and PLEX-*Bgl*II/tn43 may also be essential. In addition genes outside this region within the remaining cosmid clone (cHRIM5) may also encode products with nematocidal activity, or may enhance the nematocidal activity of genes in the smaller region (*Bgl*II-*Nru*I of cHRIM10 and PLEX-*Bgl*II/tn43).

Example 8 - Field trials

Activity of strains selected in accordance with the above methods, or from depositary institutions which include bacteria which in nature are associated symbiotically with entomopathogenic nematodes, may be further assessed in field trials as follows.

Symbiotic bacteria in the absence of their nematode host can be inoculated into one or more portions of a field which is infested with nematodes, or into containers containing unsterilised soil from such a field. The bacteria can be inoculated onto the roots of plants, or into seeds. Periodically treated and untreated areas or containers can be assayed for nematode larva, egg, or cyst counts and for the presence of the inoculated bacteria by methods well known to those skilled in the art. A reduction in the number of nematode counts in areas in which the symbiote bacteria are present indicates control of the nematodes otherwise found in the untreated areas or samples.

Annex I - amino acid sequences

SEQ ID NO:1

P0-0f

ISWFATGIPTVDALLAEFWHGDKQAFPPFTCRFTHFDPDKEQDVTLPSTEEAYWLHRA
 LQGQPLHSEVYGDDGTAQAGIPYTVMDSRPQVRLLTGLPGNSPTVWPSVIEQRTWQYERI
 ADDPQCHQOVVLNSDRYGFRETVDIAYPRRPKPAVSPYPDTLPATLFDSSYDEQQQQLR
 LTFQRQHYHHLTDTEHQVLGLPDVMRSDAWGYPAARVPREGFTLEDLLAENSLIAPGTPL
 TYLGHQRVAYTGTGTGTEEKPTRQALVAYTETAVFDELALQAFNGTLSPEALEKKLIESGY
 LSVPRFNTGAESAVVVARQGYTDYGGSEAFYRPLAQRRTTVQIGKNTLHWDTHYCAVVRM
 QDAAGLYTDAAYDYRFLTPVQITDANDNQQHITLTALGQVSSGRFWGTEEGTPQGYTPPE
 DRPFTPPSSVAEALDLKPDLPVANCVMVYAPLSWMPLAHTYQEYIAGFTWQALLDAGVVTE
 DKRVCALGFRRWVQRQGIVLNGQALADSREP VHVLT LATDRYDTPDQQLRKSVTYS DGF
 GRLLQSAVYHAPGEAWQRAADGSLITDAKGAPLVAHTATRWAVSGRTEYDGKGQPVRTYP
 PFFLNAWQYLSDD SARQDLNADTHRYDPLGREYQVRTAKGYLRONRLTPWFV VNE DENDT
 LS

SEQ ID NO:2

P1-2r

YLPQRGQCDMLLVVIGIGYLNNGQEA VIIGGIRVQTRRILHTDDRTVMGIPMEGVFANLH
 RRPLSQR TVKRLRPAVIGISLTGDPDRRFR TGI EWAWN RQITRLD

SEQ ID NO:3

P2-0f

SHLPARYGGRLTTL SRKGEMTVNRGDN LHQKTPEVTVLDNRGLTVRELRYHRHFNTPPTT
 DERITRHRFTLSGQLAHSIDPRFLDLQQT DNTVNP NMIDTALTGEVVRTRSDAGNDLI
 LNDITGRPVLAINATEVTRTWQYENDTL PGRPLSITEQPAGEAGRITERFWAGNSQAEK
 NSNLAGQCVRHYDTAGLNQTD SIALNGIPLSVTRQLLPDGT DADWQGNNEPAWNDRLAPE
 NFTTLSTADATGAVLT TTTDAAGNLQRVAYDVAGLLTGSWLRLAGGTEQVIVKSLTYS AAG
 QKLREEHNGNVVTTYTYEPETQRLVG IKT KRPQGH AQGTKVLQDLRYEYDPVGNVVKVTN
 DAEVTRFWRNOKVVPENTYVYDSLYQLVSATGREMANIVQOSTLLPTPSLIDSSTYSNYS
 RTYNYDRGDNLTQIRHSAPATGNSYTTDITVSDHSNR AVLDTLTDDPAKV DALFTAGGHQ
 IPLQFGQNLVWTPRGELLKVAPVVRD GQISDQESYRYDAASQRIIKTHVQQTANSSQAQS
 TL YLPGLERHTTINGTTVKEVLHVITIGEAGRAQVRVLHWENGKPGAISNNQMRYSYDNL
 IGSSGLEVDGDGQIIISMEEYYPYGGTAVWTARSQTEADYKTVRYS GKERDATGLYYYGYR
 YYQPWAGSWLSADPAGTIDGLNLYRMVRNNPATLDDKNG LAPGNRYVFFFIHEDRIFRL
 ASANVYRTEHNKSDIIAVVEDKALDSKLF TNSIEQFFKKPKGKAILKGSPDIKERLLNNI
 VHDLSNMQVGDQLYVNAHGHS AKPFFYS DSGYSKIIMEQLQORGANYVAKDLVNKF KLPEN
 ATIKISTCHSAEGKGAHITVTSTGTNEKMRYSSIIENKGEFSRSLAGTMENELIKLQPR

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VRGNVYGYLGATTFFYGAKEKVIHLKDGNLTTGVHEGKLSMFTKKNRFSENIFGLKVKRS
LTRTNFTGSGV

SEQ ID NO:4

p-2-9r

PAAEYVRDFTITCSVEFPASRSQLPVSRPATSYATRCRLPAASVVVSTAPVASAVLRVVKF
SGASRSFQAGSLFPCQSASVPSGSSWRVTDSGMPLSAILSVWFSPAVS

SEQ ID NO:5

P3-2r

QRALLNDIGHFAPGGTDQLIQAVIDIGVLRHHFLVAPEAGNLRIVRHFHHVPHRVVLIQ
VLQHLRPLCMSLWAFGFYANKALGLRLVGVGHHAVAVLFAQFLTRGGIRQGFHDNLLCP
ARKPOPTASQQACYVIRHTLQVTGRIGGGQYRAGGIRRAQGGEVFRCQPVVPGGFIVSLP
VCVRTIRQQIARDGQRYAVKRNTVRLVQSGGVIVTHALSGQVAVLLRLTVPCPKTLCDT
ACFASRLPCDTERASG

SEQ ID NO:6

P3-6r

SDRRQTGYAYSADHYRISGRSTVCTVRAGLMNYQCWLQHAATQLSESDSPKRDAEILLGY
VTGRSRTYLIADFDETLISSEELHQLDLSLLVRRIQGEVPVAYIIGEREFWSLPFAVSPATLI
PRPDTECLVEKALELLPDSPARIILDGTGTGAIALALASERNDCYVTGVDINSDAVMLAQ
HNAEKNAGKLAIHNVNLFQSEWFAAVGNQQQFDMIVSNPPYIDERDPHLQEGDIRFERPATA
LIAAQNGMADLQAIVGQARHFLSPNGWLLLEHGWKQGTVVRNLFLEKGYQQIATFQDYGG
NERITIGRWKNKNETHS

SEQ ID NO:7

P3-7f

ARRAVRRCGYCTGRTESRVPSVTTRCATAMITLSAAAVWRWTVTDKLSVWKNTRTGALR
CGRRGVRORLITRLCVTQARSGMQRGCIITATGITSRGRGAG

SEQ ID NO:8

P5-6r

WQNGSSSTTPRYLAGCYVWYPCSRARLSGNAKSLAPDGEWMKHTLKSKASGNTFTGRLI
FTGRPTVVITDKSGANTAALTLLNAEGEPQQGIEIRQNKYLNNRIEQDHRHVKKRIRREML
GFKSFRAQT

SEQ ID NO:9

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P5-7r

ALLFLSESRVMSLIRNAFKLLHYPVDIMAQCVRWSLTIALSLRNLEEMMAKRGIFVDHAT
IPRWVLRRLVPLLSKAERKRKKPVGSRWRMDETYIKVKGQWKYLYRSVDTDGQTDGCDYR

SEQ ID NO:10

P6-3f

VHSPSGAVAPGKFFIENFADTFPAPLPLHPFIDACIQQGFQLLPCLIAIAHSGKQAFECV
LLDRALQGSQCLQALVLPVGDVNGQTAHGFLIGYTQTHISTYNGLWLFITQGVRYRFV
RQTFVCRSLSFSEDDCTN

SEQ ID NO:11

P6-3r

RTCRERPRLM DYVLT KAAEADLRATIRHTRKQWGAQVRRYITALEQGIARLAVGQGSFK
DMSALFPALRMAHCERHYVFCLPRENAPALIVAFHERMDLLTRLADRLK

SEQ ID NO:12

P6-6r

PQTIICANVGLCITDKEKTMSRLTIDITDRQHQLKALAALQKTIKQYALERLFPGMSD
SDQAWQELKALLDTRINEGMEGKGCCKSIGEILDEELAGSDRA

SEQ ID NO:13

P7-1f

NAHFLIVSKTNVMSNQDPHNKRDSLFSAPIANLGDWSFDERVAEVFPDMVKRSIPGYSN
IISMIGMLASRFVTPGSQIYDLGCSLGAATLSIRRSINADNCRIIAIDNSPAMIERCRRH
IDSFKASTPVEVIEQNILDTDIQNASMVVLNFTLQFLHPDDROKILKKIYAGLKPGGVLV
LSEKFN FEDQKIGELLFNMHHD FKRANGYSELEVSQKRSMLENVMRTDSVDTHKSRLKEV
GFQHVEVWFQCFNFGSLLAIKGTEQ

SEQ ID NO:14

P7-9f

TMIDFGNFYQLIAKHPLNHWLDSLPAQLSHWQKTSQHGFSSWVKILENLPEIKPSHLDL
KNGVIAIHEPDL SKGEKARLHNILKILMPWRKGFPSLYDVEIDTEWRSDWKWERVLP HIS
PLEGKTVLDVGC GSGYHMRMVGEQAQLVVGIDPTQLFLCQFEAIRKLLGNNQRAHLLPL
GIEQLPELQAFD TVFSMGVLYHRRSPLDHLWQLKNQLVSDGELVLES LVIEGDENQCLIP
GERYAQMRRNVYFIPSAKMLKVWLEKCGFVDVRIVDHAATTPDEQRRT EWMKTESLVDFLD
PSDHSKTIEGY PAPIRAVLIARKP

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SEQ ID NO:15

P8-4r

SLQIDREKVGLD RYPOPIERLRQPCATCDNHCHSRHQVRFFLLKEYGAALAPISSQSAI
RYQFORHTMKKGLFAMASIFSGYCGGELFHLLTDPAHESQ

SEQ ID NO:16

P9-8r

SSFRLNDDLLTNSYSEGFLMIKLEICCYSSISCALVAQNAGADRIELSASPLEGGLTPSFG
ALQQSLQRLSIPVHPVIRPRGGDFCYNMDFEAMKNDVARIRDMPGIVFGILSENGHI
DRLMRQLMSLSGNMAVTFHRAFDNCFNPHVALEQLTELGVQRILTSGQQONAEGLTL
KELMQASRGPIIMPAGVRVSNISKFLEAGMTEVHSSAGKIVPSTMKYRKVGVMSSDDR
DVDEYSHYSVDGELVESMKGVMSLIKR

SEQ ID NO:17

P10-5r

YFGKNRRFVIYVTLMERNFYGLFNGEEMSHFSKISELQDLVADLAGFEQKLKQFEGHLGL
HFEQYSADHISLRNCESKIADRWKGFLOCGQLISESIINGRPICLFDLNQPIVLLDWKI
DCVELFPYPSQKHVYHQWEHVELVLPVPPEQLICEAKLLPQPLPDNFRMKESHPPKGKNE
RLPNPILAV

SEQ ID NO:18

P10-7f

GNTVNIQVILSEKISNALIEAGAPT DSEAHVRS AKAQFGDYQANGVMAAAKKVGIPPRQ
LAEKVVSQDLQGIASKVEIAGPGFINIFLDKAWVAANIETTLKDEKLGITPVEPOTIVI
DYSAPNVAQMHVGHRLSTIIGDAAARTLEFLGHKVIRANHVGDWGTQFGMLIAYLEKIQ
NENANDMALADLEAFYREAKKH YDEDEEFAIRARNYVVKLOGGDEYCRKMWRKLVDITMS
QNQETYNRLNVTLTEKDV MGESLYNDMLPGIVADLKQRGIAVKSDGATVVYLDEFKNKEG
EPMGVIIQKKDGGYLYTTT DIACAKYRHETLNASRVLYYIDSRQHQLMQAWAIVRKTGY
IPESMSLEHHMFGMMLGKD GKPFKTRAGGTVRLSDLLDEAIERADTLIREKNPDMPEDEL
KKVVEAVGIGAVKYADLSKSRRTDYVFDWONMLAFEGNTAPYMOYAYTRVSSIFKRADID
ENSLTLPVMLNEEREQALATRL LQFEETITTVAREGTPHVMCAXYLDLAGLFSGFYEHCP
ILNADSEELRQSR LKALLTAKTLKQGLDTLGIQTVERM

SEQ ID NO:19

P11-1r

AQVSNMHLLGDIRCGIIDNDGLRFHWGDT ELFIFQGSFYICCNPRFIKKNIDKTWACNFN
FAGNSLOIQIQLADDFFCQLSRRYSHLFGSGSHHTIRLIVTKLCFGRLTDVVSFTVGSASFNQ
RIADFF

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SEQ ID NO:20

P12-1r

HARVGVLHIRCRVAFKGOHIIIPVENIVCSTALGKICIFHRANPYRFHDEFQFVFWHIWVF
LTNEGIRTLNRFIQIGQSYCAAGTGFEWFTIFAQHHAKHVFE

SEQ ID NO:21

P12-5r

YHASFQLCRRLHTFYSLNTQSIKTLLOSFRCCQSQLOAALAQFFAIGIQDRAVLITRE
QTGQIVQVCTHNMWRTFTGDGSDRFFKLQAGCQCLLAFFIQHHRQCQAVFIDIRTFKDR

SEQ ID NO:22

P13-1f

FTLREDSMSDWTGVSTFNVILETGLDNCNIYANGLNMIGVIINITPTDDEGNFVDIDVT
LNDNIKIVDYIDGSDIDGSDGWFTYGNPNEYNTIPNSQSYSLKSENSQITQIKRYVSCS
NTSRLRTKSFSKVTTSKGVISITQNSINSSRVVINAIDATNFTDDELRTTKETRFENQ
SYTSHKSSTNSLYVHTWTIPRSLKLQNRWEDYNNGWTWAQSCYYKTGADGGSESTRWLA
AGSIFPPGNYDGLWLDNDIALSGMAHKSYNVDGTINQLSFTRIIGKGSWVYNISGLDRG
HAVIIIDQYGNKYRILFHAGYENS DPYLS SIVY

SEQ ID NO:23

P14-2f

VYIKFLKLFRRITMSDNNEFFTQANNETS AVSGGVDPRTGLYNIQITLGHIVGNGNLGPT
LPLTLSYSPLNKTDIGFGIGFNGFGLSVYDRKNSLLSLSTGENYKVIETDKTVKLQKKLD
NLRFEKDLKENCYRIIHKSGDIEVLTFGNNAFDLKVPKLLNPAGHAIYIDWNFEATQP
RLNRIYDDLDGHDIPLLNLEYQGLIKTILTFPGQKEGYRTELRLNRLQNSIHNFSIGN
ENPLTWSFGYTFIGKNGILQWITSMTAPGGLKETVNYSNNNQGHFPQSANLFLVLPYVT
LMKQVPGAGQPAIQAEYSYTSYSHNYVGGGSGNGIWNKLDNLYGLMTEYNYGSTESRRYKDK
EGHDQIVRIERTYNNYHLLTSECKQONGYIQTETAYYAIIGHNFDSQPSQFQLPKTKTE
TWRSADNSYRSEITETT FDESGNPLTKVIKDKKTOKIISPSTHWEYYPAGEVDNCPPEP
YGFTFRVKKI IQTPYDSEFKDDPEKFIQYRSLIGSQSHVTLKIEERHYSATQLLNSTLF
QYNTDKSELGRLLKQTECTKGENGKTYSVVHKFTYTKQDDTLQOSHSTHNDNFTIHRSQ
VRSRYTGRLFSDTDTKDIVTQMSYDKLGRLLTRTLNSGTFYANTLTIDYELNNLQDDNRP
PFVITTTDVNGNQLRNEFDGAGRHSVQCLKDSGDGKFTYIHTQQYDEQGRHHTSTYSYD
LTNGRQOTDPKVLHLSMSKSYDNWGQIANTHWSYGVSEKITVDPITLTATKQLQSNNSNV
QTGKEVTTYTPSQQPIQITLFEAGHLQSCHTLTRDGNDRVRKETDAIGQCTIYQYDNYN
RVIQITLPDGTIVNRKYAPFSTDITDIRVNGISLGQOTFDGLSRLTQSQDGGRVWAYT
YSAGNDQCPSTVITPDGQFIHYQYQPELDDAVLQVASNEITQQFSYNPVTGALLKAVAEG
QSLTPIYYPSGRKLMENINDMKMSYLWTLRGLENGYTDLTGTIQKISRDTHGRVTQIKD

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SSIKTTLNYYDDLNRHIGSQVTDLATGHMLTTTVEFDGLNREIGRKLCDSSGHTLDIQQSW
 LKTQQLANRIVKLNGLVLRTEQYSYDSRNRLNQYKCDGAECPTDKYGHISIVTONFTYDIY
 GNITACHTTTFADGTEDHATFKFANPTDPCQLTEVHHTHPDMPDNIRLKYDKAGRVINITD
 NHGNTENFTYDTLGRQLONGQGSVYGYDPLNRLVSQKTDTLDCELYRETMLVNEVRNGEM
 IRLRLTGETIIAQQRASKVLLTGTDSSQSVILTSKQNLSEAYSAYGKHKSTANDASIL
 GYNGERADPVSQVTHLNGYRSYDPTLMRFHTPDSLSPFGAGGINPYSYCLGDPINRSDP
 SGHLSWQAWTGIGMGIAGLLLTATGGMAIAAAGGIAAAIASTSTTALAFGALSVTSDIT
 SIVSGALEDASPKASSILGWVSMGMGAAGLAESAIGGTKLATHLGAFEDGENALLKST
 SESSRIKWGVTRSLDREIVRNEEGOVIKDHSRGYTDNFMGKGEQAILVHGDKDGFYHTE
 GNMKNGKGPYTRHTPEQLVDYLKDNINVDLTQGGDKFVHLLSCYKSSGAADKMAKYINR
 PVIAYS NKPTISQGLARIERKDFFLKSTYHSYDPRKIILGRTEKTVKPKTFRP

SEQ ID NO:24

P17-6r

LCYGHICLSGIPHRHIYIGSTYYGNRKSTVLYAAILHSVSLFYLLIAVFSASSAGYLTYG
 LSYHTISVQFLGLSHQIFLLSTYDQSLNLLLDYQYGD SGHRNLE

SEQ ID NO:25

P17-8r

SAQCIVGKVFRISMVISDIYYSTSLIIFQPDIIIRHIWMSVVYLCQLAWVSWVGKFEQSMV
 FCPICECGVTGGDIAIDIISKILCDYAMAFVCFRAFRTVTFILVQPIVGIVRVLFCTLOY
 SIQFHYISIC

SEQ ID NO:26

P18-7r

PSSLRTISLSKLLVTPHFILELSEVDLSKAFSPSSANAPRCVASLVPPLMADSANPAAPI
 PIETHPSIEDAFGEASSAPLTDIVISDVTLAPNASAVVEVEAIAAAIPPAIAIAPPV
 AMVSSNPAIPMPFVHACQLK

SEQ ID NO:27

P19-5f

AHCHIALFPCWHNPQYCOQHPDHHSNCHHQFKQEYPPSRQARENITLTQLPIKHTGIEAG
 SQTNRKQOTCMFQRANESKVHQLGQNGQRDRNFYWCEDILT

SEQ ID NO:28

P19-8f

PQSTPSSQNSRQLTPAESSQHOKOKSDHIEIMI PSEAPREYREQLHKATPARNRDVAFNP
 SVFDILRDYHWKNFSPVKAAKSSLTPHFVHQKAIPLNDQRNTSMKQSLKPEMRQKLY

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FOOEEY 42855360

SEQ ID NO:29

P20-1r

GKNCINDQGNLPDRYTQNCRPHLTDNPPYGTVTERNPRQYQHADLFQMRKLIGQLQNPSPG
 NNGPTQRQHWRIAIRSHKQCKNDHTDIEQCRSKSRHRKAVPCIKNCASQRSQRNQKDIRK
 RNSK

SEQ ID NO:30

P20-9r

NNTMNLKSLAAVSSMTMFSRVLGFIIRDARIIFGAGMATDAFFVAFKLPNLLRRIFAE
 GAFSQAFVFPILAKEYKNQOGDEATRFTIAYISGMLTLILAIVSVIGVIAAPWIIYVTAPGF
 TDTPDKFVLTRDLLRITFPYIFLISLASLAGAILNTWNRFSVPAPFAPTLNVSMIIFALF
 VAPYCNPPVLALGWAVVAGGVLQLAYQLPHLKKIGMLVLPRI SFROSAVWRVIRQMGPAI
 LGVSVGQISLIINTIFASFLVSGSVSWMYADRLMELPSGVLGVALGTILLPSLAKSFSS
 GNHEEYRKLMDWGLRLCFLALPCAVAGILAEPLTVSLFQYGHFSAFDAEMTORALIAY
 CFGLMGLIVVKVLAPGFYSRQDIKTFVKIAIATLIETQLMNLAFVGPLKHAGLALSIGLA
 ACENASMLYWQLRKRDIFTPLAGWGI FLFKLVVAIAVMVGVLAVLVWMPAWEQGNMAMR
 LLRLMGVVIAGAGSYFAVLALMGFRLKDFAHRLQ

SEQ ID NO:31

P21-7r

AIILIRDKLSRIFSRQISGEGMFGYRSASPKIRFITDRMVRLVYERDAYRLAEYYSENK
 DFLKPWEPTRDGSFYQPSGWTNRLNYIAELQRQNTFNFVLLDSDEREIMGVANFTNVVR
 GAETHSCYLGYSLAEKLOGQGLMYEALQPAIRYMORYORMHRIMANYMPHNHRSGNLLKKL
 GFEQEGYAKNYLMIDGVWQDHVLTALTDDAWGKVGL

SEQ ID NO:32

P21-8f

WCAMSLVSQARSLGKYFLLFDNLLVVLGFFVVEPLISIRFVEQLGWAALIVGFALGLRQL
 VQOGLGIFGGAIADRFQAKPMIVTGMLLRALGFALMAMAHEPWILLSCVLSGLGGTLFD
 PPRAALVIKLTTPHERGRFYSLMMQDSAGAVVGALIGSWLLQYDFNIVCWIGASIFVLA
 ALFNAWLLPAYRISTIRTPIKEGMMRVIRDRRFLYYVLTLTGYFVLSVQVMLMFPIIHE
 ITGTPTAVKWMYAIETAISLTLLYPIARWSEKHFRLEQRLMAGFLMSICMPPIGWVNQL
 HTLFGLLCLFYLGVLVTADPARETLSASLSDPRARGSYMGSRLGLALGGAIGYTGGGWLY
 DTGRDLNMPQLPWILLGLSLITIIYALHRQFNQKKIDPVMGLGRH

SEQ ID NO:33

P23-1f

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KGANMKRFFLGAALVLVGLVSGCDQFKDFSINEGLMNDYLLKKVHYQKKISIPGIANANI
 TLGDLSSQIGRODPEKIELSTQAKVOLATLLGTIQADMKI.TIKAKPVFDAEKGAIFVKGL
 EIVDYQTTPEKAAAPVKALIPYLNLSLSEFFDTHPVYVLNPEKSKAEAAASQFAKRLEIK
 PGKLVIGLTDK

SEQ ID NO: 34

P24-4r

QVALQHGRRLGTITLFDNLLGLNQVMNEFSIVCRILGTLENRAPQDPVLQPLITMIAEGK
 LKQAWPLEQDEWLDRLQONSELVMAADYHALFTGESASVAVCRSDYTDGEESEVRQFLT
 ERGMPLSDTPADQFGSLLLAVSWLEDQAAEDEIQAQITLFDYLLPWCQGFLGKVEAHAT
 SGFYRTLAIIVTREALQALRDELESE

SEQ ID NO: 35

P25-3r

DCMNIIFFHPSFNTDEWIOGQIARLPDAKVRQWVSGDQEPADYALVWQPPYEMLANRQGL
 KGIFALGAGVDAlFKQESKNPGTLLADVPLIRLEDTGMRQMQEYAITSVLHYFRMDEY
 KRYQEQRLWNPIAPHNRKEFVIGVLGAGILGRSVIGKLMEFDENVRCWSRTSKQLDSVES
 FYGKEQLGDFLSGCKVLINLLPDTPDTRGILNLSLFSQLKSGSYVINLARGAQLVEQDLL
 VAIDKGYIAGATLDVFAEEPLSNMHPFWTHPRINVTPHIAANTIPEAAMDVICENIRRMV
 QGEMPTGLVDRVRGY

SEQ ID NO: 36

P26-0f

KTSQGFTSTTCSNGNVLKICGLITPCSSLIQRTYPNNMTIGIFSKESTAKNFGMGFLYYF
 DLRVLSPPFFKAPINIFTGWQHNTNFRKSRNSTIRLCSSTPNSKOYFTTSRKCHITGAGKY
 RFSIENCFIKSG

SEQ ID NO: 37

P27-0r

YSAGCSTVLKSSLNLQCDTFNCESFVMLTLNFTSVNAKPSHIWAHYVDFDLRKKWEVDL
 EYFQFEGEVKTGQYGRMILSGMPEIRFYLSNIEVNKEFTDQVNLPQMGIITFRHQIITDE
 NNMACRVQVTVSFEPDANI PAVQAESFFKQGTQDLVESVLRKSVVETVSPKPNLQLVYV
 SDIESSTAFYKTI FNAEPI FASSRYVAF PAGGEVLFAIWSGGAKPDRAIPRFSEIGIMLP
 SGKDVDRCFEEWRKNPEIKIVQEPHTEVFGRTFLAEDPDGHIIRVCPLD

SEQ ID NO: 38

P27-8r

KGNQITMILYKGSKNYLFNQLNYDSCVLLLEVDES VN LN GWDELSRAQRLLFLMEILARYH

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FPVQGVLAQKLNISLRTLYRDIASLOAQGAIIEGEPGIGYVLRPGFVLPPLMFTQNEIE
 ALALGANWVAKRADPQLKESANNATSKIAAVIPAEKQMLEASSLLIGPAATAVQPVVEI
 QQIRQAINTRHKITLAYLDIKDIPSERTIWPFAALGYFENISIVIGWCELREEFRHFRSDR
 IMRLKIENQCYPRSRQVLLKZWRAMEKISR

SEQ ID NO:39

P27-9f

RKMTIYDLKPRFQNLRLPIVIYLYKOGITANQVTLTALFLSIFAGSLLSLFSPHLYWLL
 PVFLFIRMALNAIDGMLAREHNQKSHLGAIYNELGDVISDVALYLPFCLLPDVNSLSLLI
 ILFLTILTEFIGVLAQTIGASRRYDGPIGKSDRAFIFGAYGLIIAIFPLALGWSISLFAF
 MIILLLVTCYQRVVKALREIRLAEQSHSK

SEQ ID NO:40

P28-5f

GVMNTPQLDQRIAEHHYFTTSDNASLFYRYWPPQQANPDRAIIIFHRGHEHSGRIQHVVD
 GLDLPDVPMPFAWDARGHGKTEGPRGYSPSMGTSIRDVDEFVRFIATQYGIAMENIVVIGQ
 SVGAVLVSAAVHDYAPKIRAMILAAPAFDIKLYIPFATQGLQMQKARGIFFVNSYVKAR
 YLTHDETRIASYNSDPLITREIAVNILLDLYQTAERVVKDAAAITLPTLLFISGSDYVVN
 KKPQHQFYQQLNTPIKEKHVMDGFYHDTLGEKDRHLVFDKIRVFIERIFALPRYQHDYSQ
 EDTWSHSADEFRTLSTSLPCLCPKKLSYQLMRKVMSTHWGRTSEGVCIGLKTGFDSGSTL
 DYVYRNQFPQKGILGRILDKHYLNSIGWRGIRQRKIHIEMLRHAIRSLREQNMPVHMVD
 IAAGHGRYILDAINDFSKVDSILLRDYSEINVNQQAIEERDLTDKIRFTIGDAFNAES
 ISSITPAPTLGIVSGLYELFPDNNLLRNSLRGFADVMTENGYLVTGQPWHPQIEVIARV
 LSSHRDSQPWIMRRRTQGEMDALVEAAGFEKLYQLTDNWGIFTVSIKRVHR

SEQ ID NO:41

P28-5bf

HHNSINVLLKNIISPHQIMLLCFTVTGHNNRPIQTERSLEFTVVMSTQDVSSMSLTDSIC
 LMFCLSRGMPVDTVRQKGRAVTAHPWERRFVMLMNLSDLLPLSTASPWKISWLSARVSE
 Y

SEQ ID NO:42

P30-3f

INKYKMEHHMHSSLDSERRRLWLTGVIWLLFLAPFFFLTYGQVNQFTAQRSDVGTVMFGWE
 HNIPFWSWSIIPYWSIDLFGYISLFICTHRREQWLHGWRMTASLIACVGFLLEPLKFSF
 SRPTTEGLFGWLENQLELFDLPYNQAPSLHIILLWLLWLRYSAYVSGYWRGLLHIWSVLI
 ALSVLTWQHHRFIDVLTGFAVGVLISYLLPVSYRWRWQPNQDRYARKLFGYYLTGSALFA
 LIASLLGGSFWILLWPAVSLLMIALGYAGLGSSVFQKQPDGRMSLSARWLLAPYQLGAWL
 SYLWFRKSAFFNHTEGIILGSLPCQPVTAHSVLDITAEWHRRSDARTVNYVCQPQIDL

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LPLAPEALQSAVCTLDKLRQQGDVFEVHCTLGLSRSAMVVAWLLKQHPEYDINTVVAILR
KARPHVTFRQTHLDALSQWAKGYL

SEQ ID NO:43

P31-6f

QSCVKPDRMSRSDKHIWMPCLNGQKATYNGEHNMQPENLISKVIIATLKSURFISTLSAF
STLIATAMLIHAVNTTALNNIALYAVLLFTTLYCQYYCWRTWLDCHYFQILNSSPEKSAE
FDQTLILLIFNKLPQSRTQNDRFNGAIKLLKKATIGLILQWILFFLFLTLKYSA

SEQ ID NO:44

P32-3f

MNTRKINGIRPFSAFIDSCLESYSFFRFIRDIIAGITVGVIAIPLAMALAIGSGVAPQY
GLYTAAIAGIVIAMTGGSRYSVSGPTAAFFVILYPVSQQFGLSGLLIATLMSGVILIVMG
LARFGRLEIYIPMSVTLGFTSGIAITIAMQVQNFGLKLANIPENYIDKVVALYQALPS
LQSDTLIGLITLLVLIFWPKLGVKLPGLPALIAGTAVMGAMHLLNHDVATIGSSFSYT
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SEQ ID NO:45

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SEQ ID NO:46

P33-5f

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SEQ ID NO:47

P34-3f

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48

SEQ ID NO:48

P35-0r

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SEQ ID NO:49

P35-8r

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SEQ ID NO:50

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SEQ ID NO:51

P37-5r

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Claims:

1. The use of a bacterial strain to control a target nematode, characterised in that in nature the bacterial strain is associated symbiotically with an entomopathogenic nematode.
2. The use according to claim 1, wherein the bacterial strain from nature is directly employed to control the nematode target, or is employed to give a recombinant bacterium employed to control the nematode target, or the natural or recombinant strain is employed as a source of a nematode control agent to control the nematode target.
3. The use according to claim 1 or 2, wherein the target nematode is not the same as the nematode with which the bacterial strain is found symbiotically in nature.
4. The use according to claim 1, 2 or 3, for control of helminthiasis in a human or a domesticated animal or the control of plant pathogen nematodes.
5. The use according to any preceding claim wherein the nematode to be controlled comprises one or more of *Haemonchus*, *Trichostrongylus*, *Ostertagia*, *Nematodirus*, *Cooperia*, *Ascaris*, *Bunostomum*, *Oesophagostomum*, *Chabertia*, *Trichuris*, *Strongylus*, *Trichonema*, *Dictyocaulus*, *Capillaria*, *Heterakis*, *Toxocara*, *Ascaridia*, *Oxyuris*, *Ancylostoma*, *Uncinaria*, *Toxascaris*, *Parascaris*, *Aphelenchoides*, *Anguina*, *Bursaphelenchus*, *Criconemella*, *Meloidigyne*, *Ditylenchus*, *Globodera*, *Heliocotylenchus*, *Heterodera*, *Pratylenchus*, *Radopholus*, *Rotelynchus*, *Tylenchus*, *Trichodorus*, *Xiphenema*, and *Caenorhabditis*.

6. A composition for the control of parasitic nematodes which comprises as an effective agent a species of bacterium which is a symbiont of an entomopathogenic nematode, or an engineered bacterium, or a nematode control agent derived from a natural or engineered bacterium.
7. A composition according to claim 6, wherein the bacterial species is of the genera *Xenorhabdus* or *Photorhabdus*,
8. A composition according to claim 7, wherein the bacterial species is of the genus *Xenorhabdus*
9. A composition according to claim 8, wherein the bacterial species is of , the species *Xenorhabdus bovienii*.
10. A composition according to claim 8, wherein the bacterial species is:
Xenorhabdus bovienii strain H31 deposited with NCIMB under accession number NCIMB 40985;
Xenorhabdus bovienii strain I73 deposited with NCIMB under accession number NCIMB 40986; and
Xenorhabdus strain C42 deposited with NCIMB under accession number NCIMB 41004.
11. A composition according to any of claim 6, wherein the nematode control agent which is derived from a symbiont of an entomopathogenic nematode or from an engineered bacterium has functional activity against a nematode, and is a peptide.
12. A nucleic acid encoding a peptide of claim 11.
13. A nucleic acid according to claim 12, which nucleic acid comprises a

natural nucleotide sequence or a degeneratively equivalent sequence, or a functional variant thereof.

14. A nucleic acid according to claim 13, which is a homologous variant encoding a peptide which is a nematode control agent, the nucleic acid having 70% or more DNA sequence identity and/or the peptide having 70% or more amino acid sequence identity.
15. A nucleic acid according to claim 13, which is all or part of cosmid cHRIM5, in particular p 13-1f or p 14-2f, and variants thereof.
16. A nucleic acid according to claim 13, 14 or 15, wherein the variant has a sequence which is a derivative by way of addition, insertion, deletion or substitution of one or more nucleotides.
17. A nucleic acid according to any of claims 12 to 16, which is part of a longer sequence and the nematode control agent is expressed as a fusion protein.
18. A nucleic acid complementary to a nucleic acid according to any of claims 12 to 17.
19. A nucleic acid for use as a probe or primer having a nucleotide sequence of at least 15 nucleotides, which sequence is present in a nucleic acid according to any of claims 12 to 18.
20. A method for identifying or cloning a nucleic acid according to any of claim 12 for a nematode control agent, which method employs a nucleic acid probe according to claim 19.

21. A method according to claim 20, which comprises the steps of:
- (a) providing a preparation of nucleic acid from a bacterium,
 - (b) providing a probe,
 - (c) contacting nucleic acid in said preparation with said probe under conditions for hybridisation of probe to any said gene or homologue in said preparation, and,
 - (d) identifying said gene or homologue if present by its hybridisation with said probe.
22. A method according to claim 20, which comprises the use of two primers to amplify a nucleic acid encoding a nematode control agent, at least one of the primers having a conserved nucleotide sequence of at least 15 nucleotides.
23. A method according to claim 20, which comprising the steps of:
- (a) providing a preparation of nucleic acid from a bacterium,
 - (b) providing a pair of nucleic acid molecule primers, at least one of which is a primer,
 - (c) contacting nucleic acid in said preparation with said primers under conditions for performance of PCR,
 - (d) performing PCR and determining the presence of absence of an amplified PCR product.
24. A recombinant vector comprising a nucleic acid according to any of claims 12 to 17.
25. A host cell containing a vector according to claim 24 capable of replication.
26. A host cell according to claim 25 which is a plant cell.

27. A method for producing a transgenic plant which comprises the step of regenerating a plant from a plant cell according to claim 26.
28. A plant produced according to claim 27 which is a crop species which can be maize, cotton, soya, rice, *Brassica* species, tomato, potato, sugar beet, barley, soybean, peanut, onion, rye, wheat, corn, banana, raspberry, bean, or a decorative or other plant.
29. A method of producing a peptide nematode control agent comprising causing or allowing expression of a nucleic acid according to claim 12.
30. An antibody or fragment thereof, or a polypeptide comprising the antigen-binding domain of the antibody, capable of specifically binding a peptide of claim 11.

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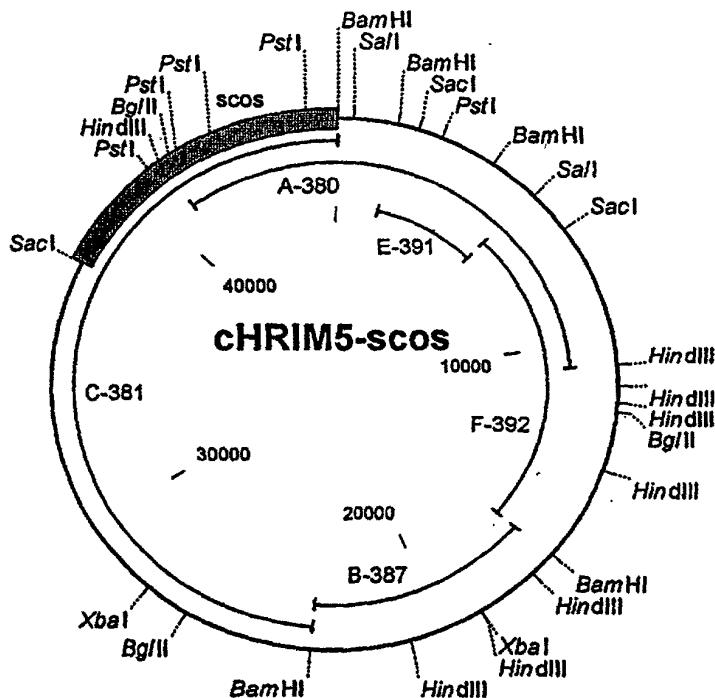

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁷ : A01N 63/00, 63/02, C12N 15/31, C12P 21/00, C07K 14/24 // (C12P 21/00, C12R 1:01)		A1	(11) International Publication Number: WO 00/42855
			(43) International Publication Date: 27 July 2000 (27.07.00)
(21) International Application Number: PCT/GB00/00219		(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).	
(22) International Filing Date: 24 January 2000 (24.01.00)			
(30) Priority Data: 9901499.5 22 January 1999 (22.01.99) GB			
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(54) Title: BIOLOGICAL CONTROL OF NEMATODES

(57) Abstract

Nematodes can be controlled through the use of bacteria associated symbiotically with an entomopathogenic nematode. The bacteria can be employed for nematode control, or engineered to a recombinant form. Control may be achieved using material such as a peptide. The peptide can be obtained from a natural or engineered nucleic acid.



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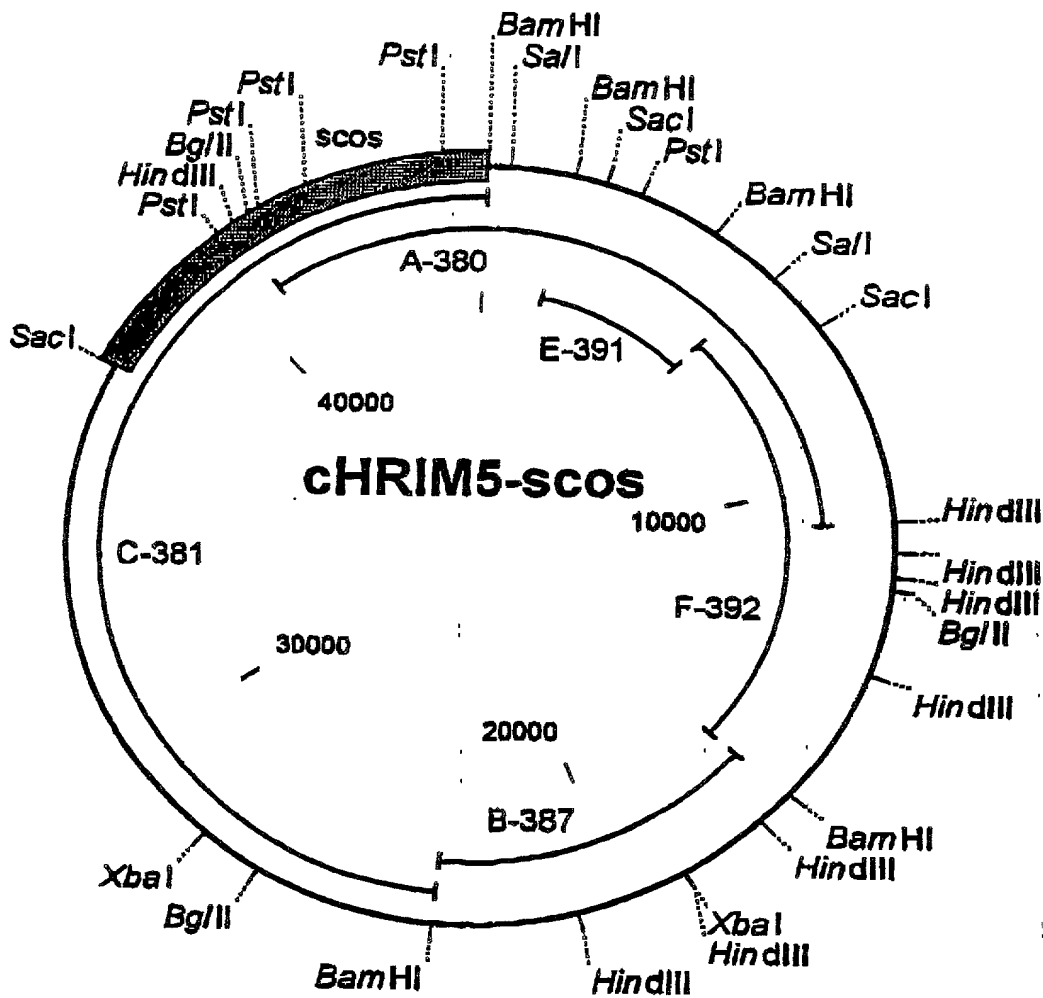


Fig. 1

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Molecule:

Sequence Data

Description:

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37544 bps DNA

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421	cacggggagac	cgtcgacatt	gcttatccgc	gccgccctaa	gcctgcccgtg	tcacctttacc
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Fig. 2

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Fig. 2(i)

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chr5ed2.seq

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8401	tcgatccaac	ccaacttttt	ctctgtcaat	ttgaagcgat	cagaaggttg	ttgggggaca
8461	atcaacgagc	ccaactttctg	ccattgggca	togaacaatt	acccgaactg	caagcctttg
8521	atacggtatt	ttcaatggga	gtgctctacc	accgcccgtc	acctcttgat	catctgtggc
8581	aactgaaaaa	tcaactgggtg	tctgatgggtg	agttagtgtc	ggaaagttaa	gtgattgagg
8641	gtgatgaaaa	tcagtgcctc	attccgggtg	aacgctatgc	acaaatgcgg	aatgtctact
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8761	gaattgtcga	tcatgcccgt	acaacacctg	atgaacagcg	ccggacagaa	tggatgaaga
8821	ccgaatcact	ggtagatttc	cttgaccctt	cagatcacag	taaaacaatt	gaaggctacc
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Fig. 2(ii)

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chr5ed2.seq

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12721	aaaaaacctg	atttttatttc	actattaata	attaatgata	attttctattt	taataaacct
12781	tggtataaaa	aatagttattt	taaaaaaaca	ttttacatta	tataaaatat	atcaatcgac
12841	tctttatttc	tttatocatt	tataaaatat	attttttacc	aaaataatat	ttaaatcata
12901	tattatattt	acatcacgtt	agatcaaaaat	aacnaattttt	tagtcgttaa	cccagattca
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13021	ggagataaat	cttcgcattt	cttcaatgaa	gatggatata	gatactgtaa	ggtagtaatt
13081	taaatgaaat	ccattaaaca	attaaatattt	aattttacatt	aagagaggat	tctatgagtg
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13201	tctacgttaa	tgggcttaac	atgattgggg	taattattaa	tatcacacco	actgatgatg
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15301	atcaaatagt	ccgtatagaa	cgcacataca	ataattacca	tctgttaact	tccgaattgt
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16801	cgctgataac	agatatccga	gtgaatggaa	tttctctggg	acagcaaacg	tttgacgggt
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Fig. 2(iii)

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chr15ed2.seq

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17581	acaaatattg	ccatagcata	gtcacacaaa	attttactta	tgatatctat	ggcaatatca
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Fig. 2(iv)

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Fig. 2(v)

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Fig. 2(vi)

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Fig. 2(vii)

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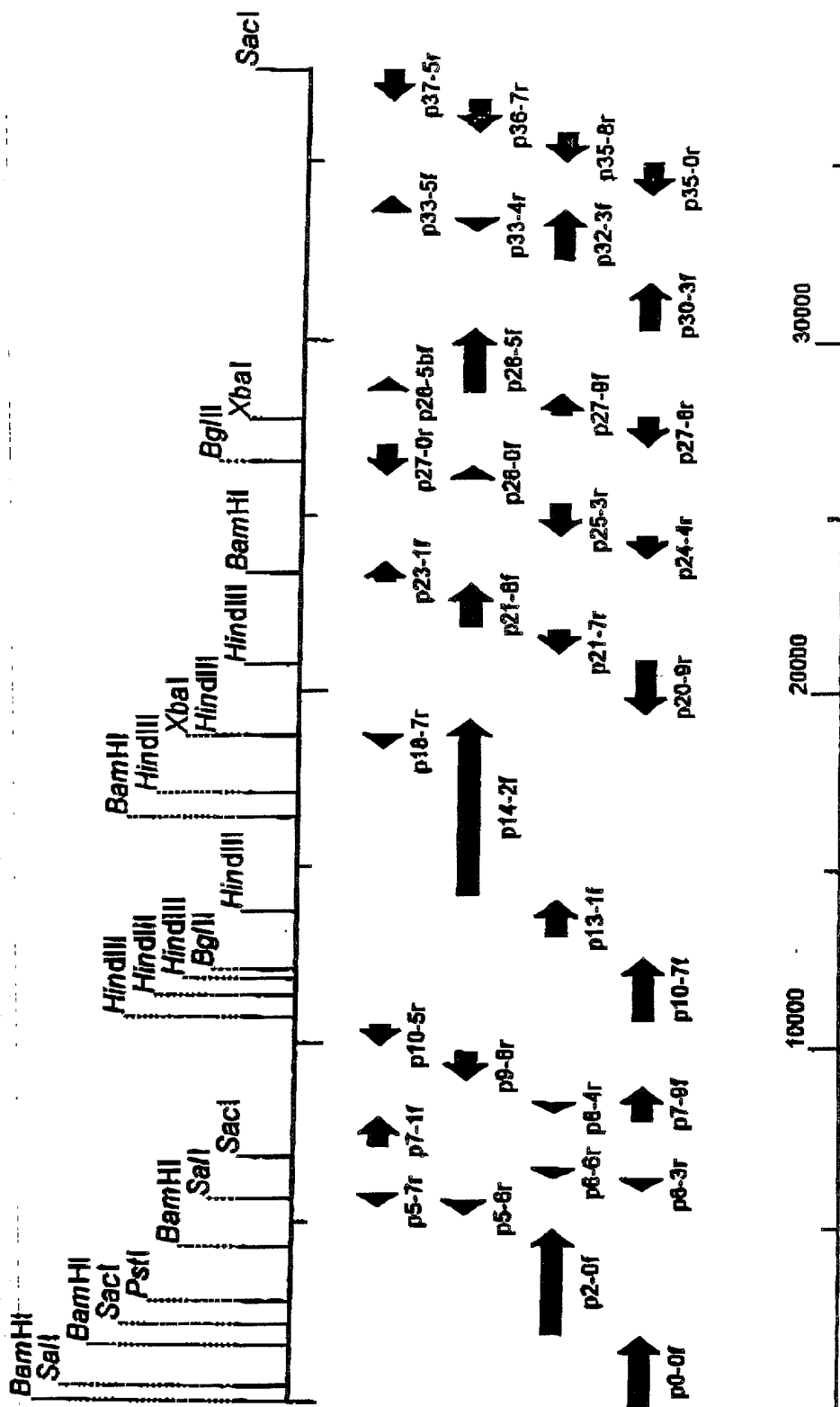
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Fig. 2(viii)

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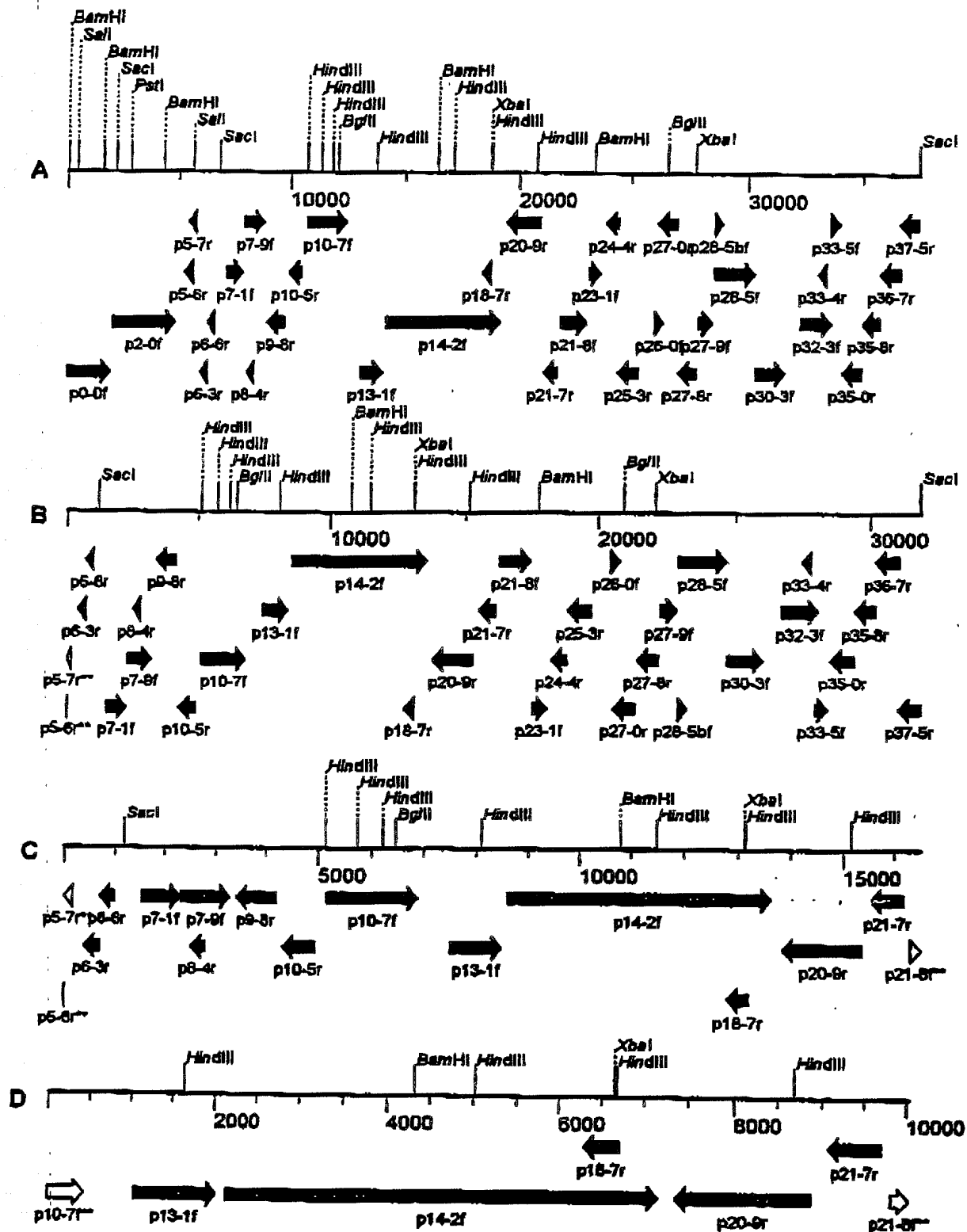


Fig. 4

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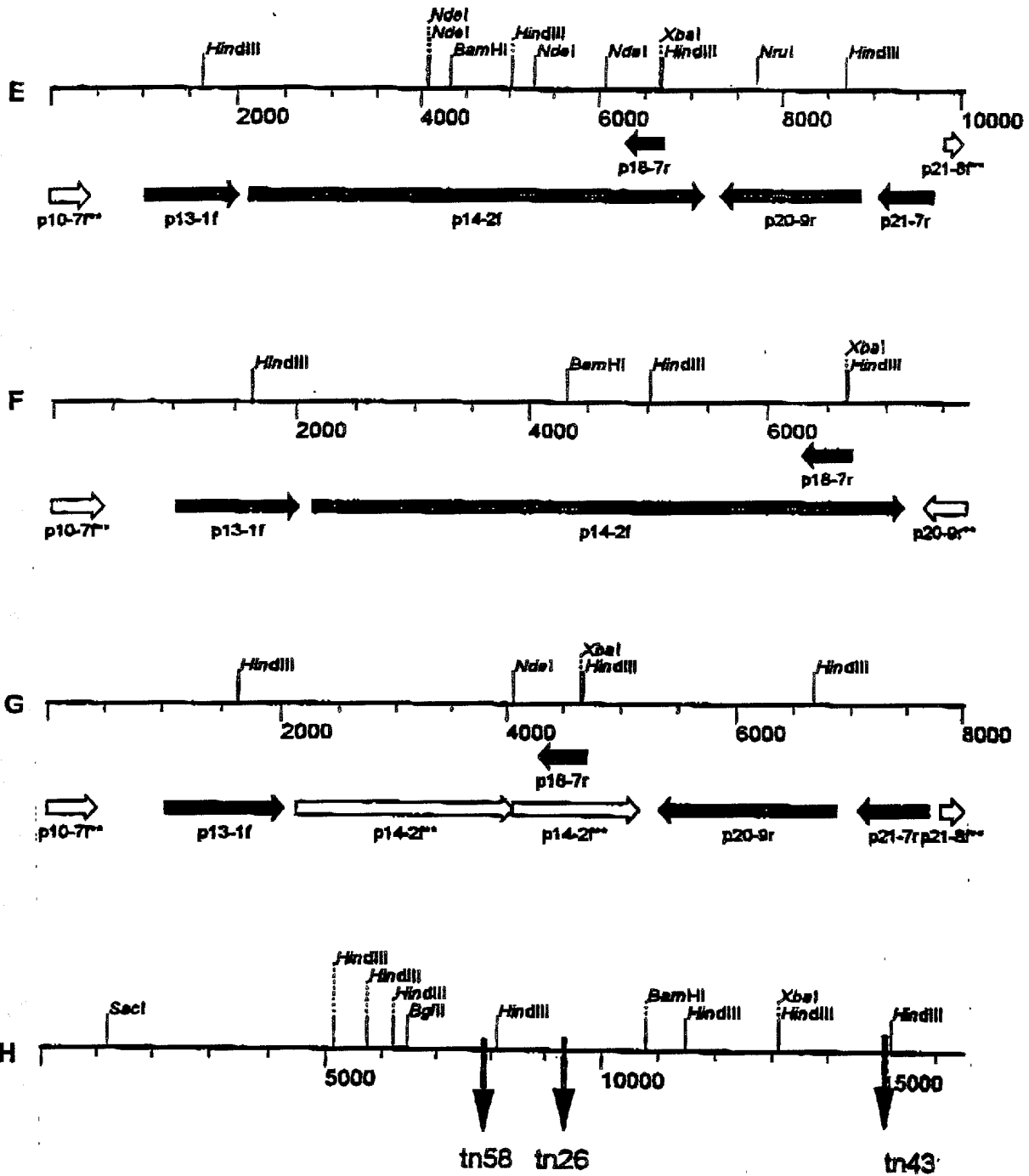


Fig. 4(cont'd)

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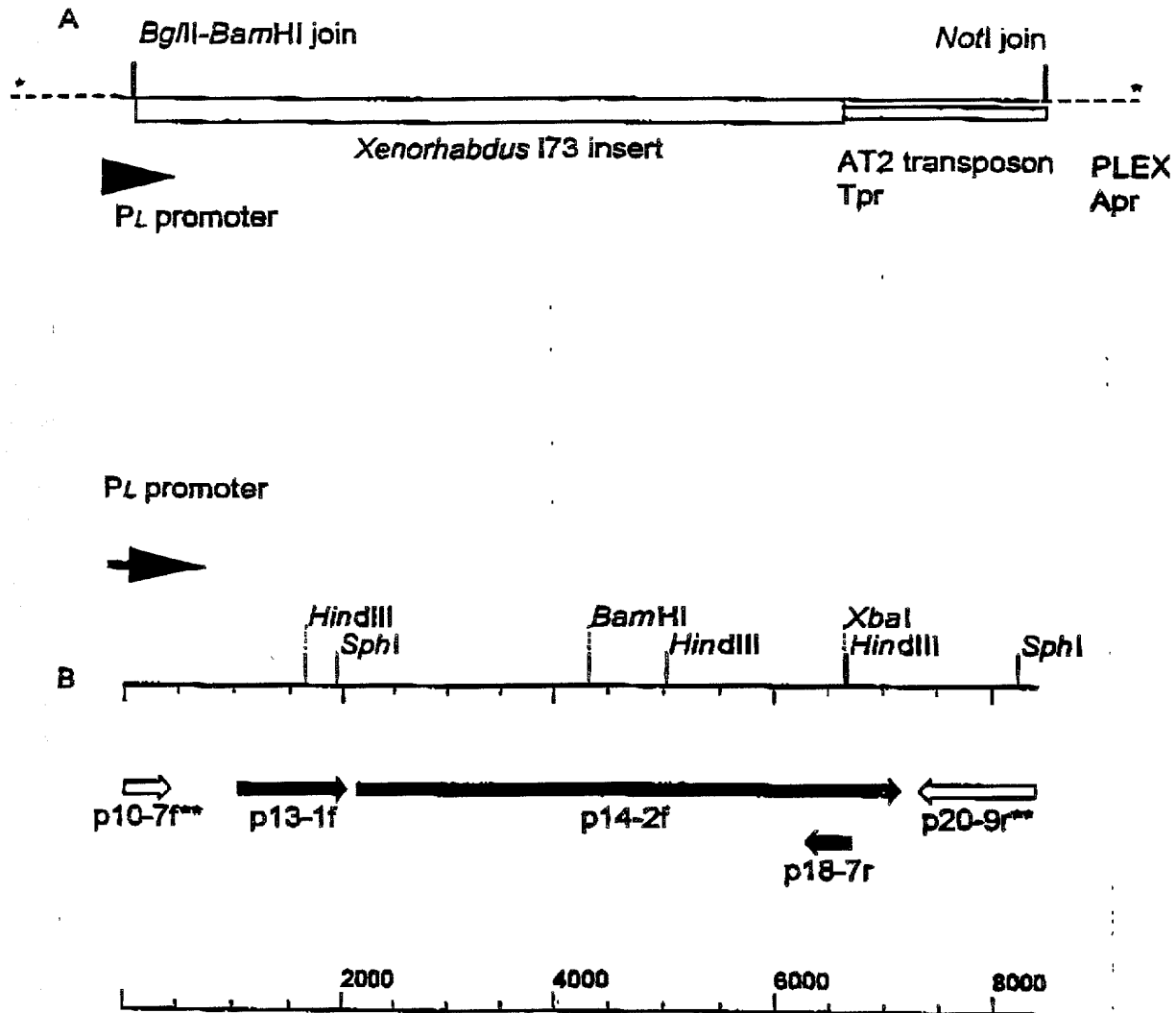


Fig. 5

COMBINED DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled BIOLOGICAL CONTROL OF NEMATODES, the specification of which:

- ☐ is attached hereto.
☐ was filed on _ as Application Serial No. _ and was amended on _____.
☒ was described and claimed in PCT International Application No. PCT/GB00/00219 filed on 24 January 2000 and as amended under PCT Article 19 on _____.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, §1.56.

I hereby claim the benefit under Title 35, United States Code, §119(e)(1) of any United States provisional application(s) listed below:

U.S. Serial No.	Filing Date	Status
-----------------	-------------	--------

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose all information I know to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56(a) which became available between the filing date of the prior application and the national or PCT international filing date of this application:

U.S. Serial No.	Filing Date	Status
PCT/GB00/00219	January 24, 2000	Pending

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

Country	Application No.	Filing Date	Priority Claimed
Great Britain	9901499.5	22 January 1999	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Yes <input type="checkbox"/> No

I hereby appoint the following attorneys and/or agents to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

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Gly	Glu	Ala	Gly	Arg	Ile	Thr	Glu	Arg	Phe	Val	Trp	Ala	Gly	Asn	Ser	
			165						170					175		
Gln	Ala	Glu	Lys	Asn	Ser	Asn	Leu	Ala	Gly	Gln	Cys	Val	Arg	His	Tyr	
			180					185					190			
Asp	Thr	Ala	Gly	Leu	Asn	Gln	Thr	Asp	Ser	Ile	Ala	Leu	Asn	Gly	Ile	
	195						200					205				
Pro	Leu	Ser	Val	Thr	Arg	Gln	Leu	Leu	Pro	Asp	Gly	Thr	Asp	Ala	Asp	

210 215 220
 Trp Gln Gly Asn Asn Glu Pro Ala Trp Asn Asp Arg Leu Ala Pro Glu
 225 230 235 240
 Asn Phe Thr Thr Leu Ser Thr Ala Asp Ala Thr Gly Ala Val Leu Thr
 245 250 255
 Thr Thr Asp Ala Ala Gly Asn Leu Gln Arg Val Ala Tyr Asp Val Ala
 260 265 270
 Gly Leu Leu Thr Gly Ser Trp Leu Arg Leu Ala Gly Gly Thr Glu Gln
 275 280 285
 Val Ile Val Lys Ser Leu Thr Tyr Ser Ala Ala Gly Gln Lys Leu Arg
 290 295 300
 Glu Glu His Gly Asn Gly Val Val Thr Thr Tyr Thr Tyr Glu Pro Glu
 305 310 315 320
 Thr Gln Arg Leu Val Gly Ile Lys Thr Lys Arg Pro Gln Gly His Ala
 325 330 335
 Gln Gly Thr Lys Val Leu Gln Asp Leu Arg Tyr Glu Tyr Asp Pro Val
 340 345 350
 Gly Asn Val Val Lys Val Thr Asn Asp Ala Glu Val Thr Arg Phe Trp
 355 360 365
 Arg Asn Gln Lys Val Val Pro Glu Asn Thr Tyr Val Tyr Asp Ser Leu
 370 375 380
 Tyr Gln Leu Val Ser Ala Thr Gly Arg Glu Met Ala Asn Ile Val Gln
 385 390 395 400
 Gln Ser Thr Leu Leu Pro Thr Pro Ser Leu Ile Asp Ser Ser Thr Tyr
 405 410 415
 Ser Asn Tyr Ser Arg Thr Tyr Asn Tyr Asp Arg Gly Asp Asn Leu Thr
 420 425 430
 Gln Ile Arg His Ser Ala Pro Ala Thr Gly Asn Ser Tyr Thr Thr Asp
 435 440 445
 Ile Thr Val Ser Asp His Ser Asn Arg Ala Val Leu Asp Thr Leu Thr
 450 455 460
 Asp Asp Pro Ala Lys Val Asp Ala Leu Phe Thr Ala Gly Gly His Gln
 465 470 475 480
 Ile Pro Leu Gln Pro Gly Gln Asn Leu Val Trp Thr Pro Arg Gly Glu
 485 490 495
 Leu Leu Lys Val Ala Pro Val Val Arg Asp Gly Gln Ile Ser Asp Gln
 500 505 510
 Glu Ser Tyr Arg Tyr Asp Ala Ala Ser Gln Arg Ile Ile Lys Thr His
 515 520 525
 Val Gln Gln Thr Ala Asn Ser Ser Gln Ala Gln Ser Thr Leu Tyr Leu
 530 535 540
 Pro Gly Leu Glu Arg His Thr Thr Ile Asn Gly Thr Thr Val Lys Glu
 545 550 555 560
 Val Leu His Val Ile Thr Ile Gly Glu Ala Gly Arg Ala Gln Val Arg
 565 570 575
 Val Leu His Trp Glu Asn Gly Lys Pro Gly Ala Ile Ser Asn Asn Gln
 580 585 590
 Met Arg Tyr Ser Tyr Asp Asn Leu Ile Gly Ser Ser Gly Leu Glu Val
 595 600 605
 Asp Gly Asp Gly Gln Ile Ile Ser Met Glu Glu Tyr Tyr Pro Tyr Gly
 610 615 620
 Gly Thr Ala Val Trp Thr Ala Arg Ser Gln Thr Glu Ala Asp Tyr Lys
 625 630 635 640
 Thr Val Arg Tyr Ser Gly Lys Glu Arg Asp Ala Thr Gly Leu Tyr Tyr
 645 650 655
 Tyr Gly Tyr Arg Tyr Tyr Gln Pro Trp Ala Gly Ser Trp Leu Ser Ala
 660 665 670

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<210> 4
<211> 108
<212> PRT
<213> Xenorhabdus bovienii
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<400> 4															
Pro	Ala	Ala	Glu	Tyr	Val	Arg	Asp	Phe	Thr	Ile	Thr	Cys	Ser	Val	Pro
1				5					10					15	
Pro	Ala	Ser	Arg	Ser	Gln	Leu	Pro	Val	Ser	Arg	Pro	Ala	Thr	Ser	Tyr
			20					25					30		
Ala	Thr	Arg	Cys	Arg	Leu	Pro	Ala	Ala	Ser	Val	Val	Val	Ser	Thr	Ala
		35				40						45			
Pro	Val	Ala	Ser	Ala	Val	Leu	Arg	Val	Val	Lys	Phe	Ser	Gly	Ala	Ser
	50					55					60				
Arg	Ser	Phe	Gln	Ala	Gly	Ser	Leu	Phe	Pro	Cys	Gln	Ser	Ala	Ser	Val
65					70					75					80
Pro	Ser	Gly	Ser	Ser	Trp	Arg	Val	Thr	Asp	Ser	Gly	Met	Pro	Leu	Ser
				85					90					95	

Ala Ile Leu Ser Val Trp Phe Ser Pro Ala Val Ser
100 105

<210> 5
<211> 256
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<213> Xenorhabdus bovienii

<400> 5
Gln Arg Ala Leu Leu Asn Asp Ile Gly His Phe Ala Pro Gly Gly Thr
1 5 10 15
Asp Gln Leu Ile Gln Ala Val Ile Asp Ile Gly Val Leu Arg His His
20 25 30
Phe Leu Val Ala Pro Glu Ala Gly Asn Leu Arg Ile Val Arg His Phe
35 40 45
His His Val Pro His Arg Val Val Leu Ile Ala Gln Val Leu Gln His
50 55 60
Leu Arg Pro Leu Cys Met Ser Leu Trp Ala Phe Gly Phe Tyr Ala Asn
65 70 75 80
Lys Ala Leu Gly Leu Arg Leu Val Gly Val Gly Gly His His Ala Val
85 90 95
Ala Val Leu Phe Ala Gln Phe Leu Thr Arg Gly Gly Ile Arg Gln Gly
100 105 110
Phe His Asp Asn Leu Leu Cys Pro Ala Arg Lys Pro Gln Pro Thr Ala
115 120 125
Ser Gln Gln Ala Cys Tyr Val Ile Arg His Thr Leu Gln Val Thr Gly
130 135 140
Arg Ile Gly Gly Gly Gln Tyr Arg Ala Gly Gly Ile Arg Arg Ala Gln
145 150 155 160
Gly Gly Glu Val Phe Arg Cys Gln Pro Val Val Pro Gly Gly Phe Ile
165 170 175
Val Ser Leu Pro Val Cys Val Arg Thr Ile Arg Gln Gln Leu Ala Arg
180 185 190
Asp Gly Gln Arg Tyr Ala Val Lys Arg Asn Thr Val Arg Leu Val Gln
195 200 205
Ser Gly Gly Val Ile Val Thr His Ala Leu Ser Gly Gln Val Ala Val
210 215 220
Leu Leu Arg Leu Thr Val Pro Cys Pro Asp Lys Thr Leu Cys Asp Thr
225 230 235 240
Ala Cys Phe Ala Ser Arg Leu Phe Cys Asp Thr Glu Arg Ala Ser Gly
245 250 255

<210> 6
<211> 316
<212> PRT
<213> Xenorhabdus bovienii

<400> 6
Ser Asp Arg Arg Gln Thr Gly Tyr Ala Tyr Ser Ala Asp His Tyr Arg
1 5 10 15
Ile Ser Gly Arg Ser Thr Val Cys Thr Val Arg Ala Gly Leu Met Asn
20 25 30
Tyr Gln Cys Trp Leu Gln His Ala Ala Thr Gln Leu Ser Glu Ser Asp
35 40 45
Ser Pro Lys Arg Asp Ala Glu Ile Leu Leu Gly Tyr Val Thr Gly Arg
50 55 60
Ser Arg Thr Tyr Leu Ile Ala Phe Asp Glu Thr Leu Ile Ser Ser Glu

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65 70 75 80
 Glu Leu His Gln Leu Asp Ser Leu Leu Val Arg Arg Ile Gln Gly Glu
 85 90 95
 Pro Val Ala Tyr Ile Ile Gly Glu Arg Glu Phe Trp Ser Leu Pro Phe
 100 105 110
 Ala Val Ser Pro Ala Thr Leu Ile Pro Arg Pro Asp Thr Glu Cys Leu
 115 120 125
 Val Glu Lys Ala Leu Glu Leu Leu Pro Asp Ser Pro Ala Arg Ile Leu
 130 135 140
 Asp Leu Gly Thr Gly Thr Gly Ala Ile Ala Leu Ala Leu Ala Ser Glu
 145 150 155 160
 Arg Asn Asp Cys Tyr Val Thr Gly Val Asp Ile Asn Ser Asp Ala Val
 165 170 175
 Met Leu Ala Gln His Asn Ala Glu Lys Asn Ala Gly Lys Leu Ala Ile
 180 185 190
 His Asn Val Asn Phe Leu Gln Ser Glu Trp Phe Ala Ala Val Gly Asn
 195 200 205
 Gln Gln Phe Asp Met Ile Val Ser Asn Pro Pro Tyr Ile Asp Glu Arg
 210 215 220
 Asp Pro His Leu Gln Glu Gly Asp Ile Arg Phe Glu Pro Ala Thr Ala
 225 230 235 240
 Leu Ile Ala Ala Gln Asn Gly Met Ala Asp Leu Gln Ala Ile Val Gly
 245 250 255
 Gln Ala Arg His Phe Leu Ser Pro Asn Gly Trp Leu Leu Leu Glu His
 260 265 270
 Gly Trp Lys Gln Gly Thr Val Val Arg Asn Leu Phe Leu Glu Lys Gly
 275 280 285
 Tyr Gln Gln Ile Ala Thr Phe Gln Asp Tyr Gly Gly Asn Glu Arg Ile
 290 295 300
 Thr Ile Gly Arg Trp Asn Lys Asn Glu Thr His Ser
 305 310 315

<210> 7

<211> 102

<212> PRT

<213> Xenorhabdus bovienii

<400> 7

Ala Arg Arg Ala Val Arg Arg Cys Gly Tyr Cys Thr Gly Arg Thr Glu
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 Ser Arg Val Pro Ser Val Thr Thr Arg Cys Ala Thr Ala Met Ile Thr
 20 25 30
 Leu Ser Ala Ala Val Trp Arg Trp Thr Val Thr Asp Lys Leu Ser
 35 40 45
 Val Trp Lys Asn Thr Thr Arg Thr Gly Ala Leu Arg Cys Gly Arg Arg
 50 55 60
 Gly Val Arg Gln Arg Leu Ile Thr Arg Leu Cys Val Thr Gln Ala Arg
 65 70 75 80
 Ser Gly Met Gln Arg Gly Cys Ile Ile Thr Ala Thr Gly Ile Thr Ser
 85 90 95
 Arg Gly Arg Gly Ala Gly
 100

<210> 8

<211> 130

<212> PRT

<213> Xenorhabdus bovienii

12969660

[illegible]

<211> 119

<212> PRT

<213> Xenorhabdus bovienii

[illegible]

<210> 10

<211> 138

<212> PRT

<213> Xenorhabdus bovienii

Val	His	Ser	Pro	Ser	Gly	Ala	Val	Ala	Pro	Gly	Lys	Phe	Phe	Ile	Glu
1				5					10					15	
Asn	Phe	Ala	Asp	Thr	Phe	Pro	Ala	Pro	Leu	Pro	Leu	His	Pro	Phe	Ile
			20					25					30		
Asp	Ala	Cys	Ile	Gln	Gln	Gly	Phe	Gln	Leu	Leu	Pro	Cys	Leu	Ile	Ala
		35					40					45			
Ile	Ala	His	Ser	Gly	Lys	Gln	Ala	Phe	Glu	Cys	Val	Leu	Leu	Asp	Arg

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<210> 11
<211> 110
<212> PRT
<213> Xenorhabdus bovienii
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<210> 12
<211> 103
<212> PRT
<213> Xenorhabdus bovienii
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<210> 13
<211> 265
<212> PRT
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<213> Xenorhabdus bovienii

<400> 13

Asn Ala His Phe Leu Ile Val Ser Lys Thr Asn Val Val Met Ser Asn
 1 5 10 15
 Gln Asp Pro His Asn Lys Arg Asp Ser Leu Phe Ser Ala Pro Ile Ala
 20 25 30
 Asn Leu Gly Asp Trp Ser Phe Asp Glu Arg Val Ala Glu Val Phe Pro
 35 40 45
 Asp Met Val Lys Arg Ser Ile Pro Gly Tyr Ser Asn Ile Ile Ser Met
 50 55 60
 Ile Gly Met Leu Ala Ser Arg Phe Val Thr Pro Gly Ser Gln Ile Tyr
 65 70 75 80
 Asp Leu Gly Cys Ser Leu Gly Ala Ala Thr Leu Ser Ile Arg Arg Ser
 85 90 95
 Ile Asn Ala Asp Asn Cys Arg Ile Ile Ala Ile Asp Asn Ser Pro Ala
 100 105 110
 Met Ile Glu Arg Cys Arg Arg His Ile Asp Ser Phe Lys Ala Ser Thr
 115 120 125
 Pro Val Glu Val Ile Glu Gln Asn Ile Leu Asp Thr Asp Ile Gln Asn
 130 135 140
 Ala Ser Met Val Val Leu Asn Phe Thr Leu Gln Phe Leu His Pro Asp
 145 150 155 160
 Asp Arg Gln Lys Ile Leu Lys Lys Ile Tyr Ala Gly Leu Lys Pro Gly
 165 170 175
 Gly Val Leu Val Leu Ser Glu Lys Phe Asn Phe Glu Asp Gln Lys Ile
 180 185 190
 Gly Glu Leu Leu Phe Asn Met His Asp Phe Lys Arg Ala Asn Gly
 195 200 205
 Tyr Ser Glu Leu Glu Val Ser Gln Lys Arg Ser Met Leu Glu Asn Val
 210 215 220
 Met Arg Thr Asp Ser Val Asp Thr His Lys Ser Arg Leu Lys Glu Val
 225 230 235 240
 Gly Phe Gln His Val Glu Val Trp Phe Gln Cys Phe Asn Phe Gly Ser
 245 250 255
 Leu Leu Ala Ile Lys Gly Thr Glu Gln
 260 265

<210> 14

<211> 324

<212> PRT

<213> Xenorhabdus bovienii

<400> 14

Thr Met Ile Asp Phe Gly Asn Phe Tyr Gln Leu Ile Ala Lys His Pro
 1 5 10 15
 Leu Asn His Trp Leu Asp Ser Leu Pro Ala Gln Leu Ser His Trp Gln
 20 25 30
 Lys Thr Ser Gln His Gly Gln Phe Ser Ser Trp Val Lys Ile Leu Glu
 35 40 45
 Asn Leu Pro Glu Ile Lys Pro Ser His Leu Asp Leu Lys Asn Gly Val
 50 55 60
 Ile Ala Ile His Glu Pro Asp Leu Ser Lys Gly Glu Lys Ala Arg Leu
 65 70 75 80
 His Asn Ile Leu Lys Ile Leu Met Pro Trp Arg Lys Gly Pro Phe Ser
 85 90 95
 Leu Tyr Asp Val Glu Ile Asp Thr Glu Trp Arg Ser Asp Trp Lys Trp

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<210> 15
<211> 100
<212> PRT
<213> Xenorhabdus bovienii
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<210> 16
<211> 267
<212> PRT
<213> Xenorhabdus bovienii
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<400> 16

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<210> 17
<211> 189
<212> PRT
<213> Xenorhabdus bovienii
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Tyr 1	Phe	Gly	Lys	Asn 5	Arg	Arg	Phe	Val	Ile 10	Tyr	Val	Thr	Leu	Met 15	Glu
Arg	Asn	Phe	Tyr 20	Gly	Leu	Phe	Asn	Gly 25	Glu	Glu	Met	Ser	His 30	Phe	Ser
Lys	Ile	Ser 35	Glu	Leu	Gln	Asp	Leu 40	Val	Ala	Asp	Leu	Ala 45	Gly	Phe	Glu
Gln	Lys 50	Leu	Lys	Gln	Phe	Glu 55	Gly	His	Leu	Gly	Leu 60	His	Phe	Glu	Gln
Tyr 65	Ser	Ala	Asp	His 70	Ile	Ser	Leu	Arg	Cys 75	Asn	Glu	Ser	Lys 80	Ile	Ala
Asp	Arg	Trp	Arg 85	Lys	Gly	Phe	Leu	Gln	Cys 90	Gly	Gln	Leu	Ile 95	Ser	Glu
Ser	Ile	Ile	Asn 100	Gly	Arg	Pro	Ile	Cys 105	Leu	Phe	Asp	Leu 110	Asn	Gln	Pro
Ile	Val	Leu 115	Leu	Asp	Trp	Lys 120	Ile	Asp	Cys	Val	Glu	Leu 125	Pro	Tyr	Pro

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<210> 18
<211> 579
<212> PRT
<213> Xenorhabdus bovienii
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<400>	18														
Gly	Asn	Thr	Val	Asn	Ile	Gln	Val	Ile	Leu	Ser	Glu	Lys	Ile	Ser	Asn
1				5					10					15	
Ala	Leu	Ile	Glu	Ala	Gly	Ala	Pro	Thr	Asp	Ser	Glu	Ala	His	Val	Arg
			20					25					30		
Gln	Ser	Ala	Lys	Ala	Gln	Phe	Gly	Asp	Tyr	Gln	Ala	Asn	Gly	Val	Met
		35				40						45			
Ala	Ala	Ala	Lys	Lys	Val	Gly	Ile	Pro	Pro	Arg	Gln	Leu	Ala	Glu	Lys
	50					55					60				
Val	Val	Ser	Gln	Leu	Asp	Leu	Gln	Gly	Ile	Ala	Ser	Lys	Val	Glu	Ile
65					70					75					80
Ala	Gly	Pro	Gly	Phe	Ile	Asn	Ile	Phe	Leu	Asp	Lys	Ala	Trp	Val	Ala
				85					90					95	
Ala	Asn	Ile	Glu	Thr	Thr	Leu	Lys	Asp	Glu	Lys	Leu	Gly	Ile	Thr	Pro
			100					105					110		
Val	Glu	Pro	Gln	Thr	Ile	Val	Ile	Asp	Tyr	Ser	Ala	Pro	Asn	Val	Ala
		115					120					125			
Lys	Gln	Met	His	Val	Gly	His	Leu	Arg	Ser	Thr	Ile	Ile	Gly	Asp	Ala
	130					135					140				
Ala	Ala	Arg	Thr	Leu	Glu	Phe	Leu	Gly	His	Lys	Val	Ile	Arg	Ala	Asn
145					150					155					160
His	Val	Gly	Asp	Trp	Gly	Thr	Gln	Phe	Gly	Met	Leu	Ile	Ala	Tyr	Leu
				165					170					175	
Glu	Lys	Ile	Gln	Asn	Glu	Asn	Ala	Asn	Asp	Met	Ala	Leu	Ala	Asp	Leu
			180					185					190		
Glu	Ala	Phe	Tyr	Arg	Glu	Ala	Lys	Lys	His	Tyr	Asp	Glu	Asp	Glu	Glu
		195					200					205			
Phe	Ala	Ile	Arg	Ala	Arg	Asn	Tyr	Val	Val	Lys	Leu	Gln	Gly	Gly	Asp
	210					215					220				
Glu	Tyr	Cys	Arg	Lys	Met	Trp	Arg	Lys	Leu	Val	Asp	Ile	Thr	Met	Ser
225					230					235					240
Gln	Asn	Gln	Glu	Thr	Tyr	Asn	Arg	Leu	Asn	Val	Thr	Leu	Thr	Glu	Lys
				245					250					255	
Asp	Val	Met	Gly	Glu	Ser	Leu	Tyr	Asn	Asp	Met	Leu	Pro	Gly	Ile	Val
			260					265					270		
Ala	Asp	Leu	Lys	Gln	Arg	Gly	Ile	Ala	Val	Lys	Ser	Asp	Gly	Ala	Thr
		275					280					285			
Val	Val	Tyr	Leu	Asp	Glu	Phe	Lys	Asn	Lys	Glu	Gly	Glu	Pro	Met	Gly
	290					295					300				
Val	Ile	Ile	Gln	Lys	Lys	Asp	Gly	Gly	Tyr	Leu	Tyr	Thr	Thr	Thr	Asp
305					310					315					320
Ile	Ala	Cys	Ala	Lys	Tyr	Arg	His	Glu	Thr	Leu	Asn	Ala	Ser	Arg	Val
				325											

Leu Tyr Tyr Ile Asp Ser Arg Gln His Gln His Leu Met Gln Ala Trp
 340 345 350
 Ala Ile Val Arg Lys Thr Gly Tyr Ile Pro Glu Ser Met Ser Leu Glu
 355 360 365
 His His Met Phe Gly Met Met Leu Gly Lys Asp Gly Lys Pro Phe Lys
 370 375 380
 Thr Arg Ala Gly Gly Thr Val Arg Leu Ser Asp Leu Leu Asp Glu Ala
 385 390 395 400
 Ile Glu Arg Ala Asp Thr Leu Ile Arg Glu Lys Asn Pro Asp Met Pro
 405 410 415
 Glu Asp Glu Leu Lys Lys Val Val Glu Ala Val Gly Ile Gly Ala Val
 420 425 430
 Lys Tyr Ala Asp Leu Ser Lys Ser Arg Thr Thr Asp Tyr Val Phe Asp
 435 440 445
 Trp Asp Asn Met Leu Ala Phe Glu Gly Asn Thr Ala Pro Tyr Met Gln
 450 455 460
 Tyr Ala Tyr Thr Arg Val Ser Ser Ile Phe Lys Arg Ala Asp Ile Asp
 465 470 475 480
 Glu Asn Ser Leu Thr Leu Pro Val Met Leu Asn Glu Glu Arg Glu Gln
 485 490 495
 Ala Leu Ala Thr Arg Leu Leu Gln Phe Glu Glu Thr Ile Thr Thr Val
 500 505 510
 Ala Arg Glu Gly Thr Pro His Val Met Cys Ala Tyr Leu Tyr Asp Leu
 515 520 525
 Ala Gly Leu Phe Ser Gly Phe Tyr Glu His Cys Pro Ile Leu Asn Ala
 530 535 540
 Asp Ser Glu Glu Leu Arg Gln Ser Arg Leu Lys Leu Ala Leu Leu Thr
 545 550 555 560
 Ala Lys Thr Leu Lys Gln Gly Leu Asp Thr Leu Gly Ile Gln Thr Val
 565 570 575
 Glu Arg Met

<210> 19
 <211> 126
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 19
 Ala Gln Val Ser Asn Met His Leu Leu Gly Asp Ile Arg Cys Gly Ile
 1 5 10 15
 Ile Asp Asn Asp Gly Leu Arg Phe His Trp Gly Asp Thr Glu Leu Phe
 20 25 30
 Ile Phe Gln Gly Ser Phe Tyr Ile Cys Cys Asn Pro Arg Phe Ile Lys
 35 40 45
 Lys Asn Ile Asp Lys Thr Trp Ala Cys Asn Phe Asn Phe Ala Gly Asn
 50 55 60
 Ser Leu Gln Ile Gln Leu Ala Asp Asp Phe Phe Cys Gln Leu Ser Arg
 65 70 75 80
 Arg Tyr Ser His Leu Phe Ser Gly Ser His His Thr Ile Arg Leu Ile
 85 90 95
 Val Thr Lys Leu Cys Phe Gly Arg Leu Thr Asp Val Ser Phe Thr Val
 100 105 110
 Gly Trp Ser Ala Ser Phe Asn Gln Arg Ile Ala Asp Phe Phe
 115 120 125

<210> 20

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<400> 22
Phe Thr Leu Arg Glu Asp Ser Met Ser Asp Trp Thr Gly Val Ser Thr
 1          5          10          15
Phe Asn Val Ile Leu Glu Thr Gly Leu Asp Asn Cys Asn Ile Tyr Ala
          20          25          30
Asn Gly Leu Asn Met Ile Gly Val Ile Ile Asn Ile Thr Pro Thr Asp
      35          40          45
Asp Glu Gly Asn Phe Val Asp Ile Asp Asp Val Thr Leu Asn Asp Asn
    50          55          60

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Ile Lys Ile Val Asp Tyr Ile Asp Gly Ser Asp Ile Asp Gly Ser Asp
 65 70 75 80
 Gly Trp Phe Tyr Thr Gly Asn Pro Asn Glu Tyr Asn Thr Ile Pro Asn
 85 90 95
 Ser Gln Ser Tyr Ser Leu Leu Lys Ser Glu Asn Ser Gln Ile Thr Gln
 100 105 110
 Ile Lys Arg Tyr Val Ser Cys Ser Asn Thr Ser Arg Leu Arg Thr Lys
 115 120 125
 Ser Phe Ser Ala Lys Val Thr Thr Thr Ser Gly Lys Val Ile Ser Ile
 130 135 140
 Thr Gln Asn Ser Ile Asn Ser Ser Arg Val Val Ile Asn Ala Ile Asp
 145 150 155 160
 Ala Thr Asn Phe Thr Asp Asp Glu Leu Arg Thr Thr Lys Glu Thr Arg
 165 170 175
 Phe Glu Asn Gln Ser Tyr Thr Ser His Lys Ser Ser Thr Asn Ser Leu
 180 185 190
 Tyr Val His Thr Trp Thr Ile Pro Arg Ser Leu Lys Leu Gln Asn Trp
 195 200 205
 Arg Trp Glu Asp Tyr Asn Asn Gly Trp Thr Trp Ala Gln Ser Cys Tyr
 210 215 220
 Tyr Lys Thr Gly Ala Asp Gly Gly Ser Glu Ser Thr Arg Trp Leu Ala
 225 230 235 240
 Ala Gly Ser Ile Phe Pro Pro Gly Asn Tyr Asp Gly Leu Trp Leu Asp
 245 250 255
 Asn Asp Ile Ala Leu Ser Gly Met Ala His Lys Ser Tyr Asn Val Asp
 260 265 270
 Thr Gly Ile Asn Gln Leu Ser Phe Thr Arg Ile Ile Gly Lys Gly Phe
 275 280 285
 Ser Trp Val Tyr Asn Ile Ser Gly Leu Asp Arg Gly His Ala Val Ile
 290 295 300
 Ile Ile Asp Gln Tyr Gly Asn Lys Tyr Arg Ile Leu Phe His Ala Gly
 305 310 315 320
 Tyr Glu Asn Ser Asp Pro Tyr Leu Ser Ser Ser Ile Val Tyr
 325 330

<210> 23
 <211> 1673
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 23
 Val Tyr Ile Lys Phe Leu Lys Leu Phe Arg Arg Ile Thr Met Ser Asp
 1 5 10 15
 Asn Asn Glu Phe Phe Thr Gln Ala Asn Asn Phe Thr Ser Ala Val Ser
 20 25 30
 Gly Gly Val Asp Pro Arg Thr Gly Leu Tyr Asn Ile Gln Ile Thr Leu
 35 40 45
 Gly His Ile Val Gly Asn Gly Asn Leu Gly Pro Thr Leu Pro Leu Thr
 50 55 60
 Leu Ser Tyr Ser Pro Leu Asn Lys Thr Asp Ile Gly Phe Gly Ile Gly
 65 70 75 80
 Phe Asn Phe Gly Leu Ser Val Tyr Asp Arg Lys Asn Ser Leu Leu Ser
 85 90 95
 Leu Ser Thr Gly Glu Asn Tyr Lys Val Ile Glu Thr Asp Lys Thr Val
 100 105 110
 Lys Leu Gln Gln Lys Lys Leu Asp Asn Leu Arg Phe Glu Lys Asp Leu
 115 120 125

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Lys	Glu	Asn	Cys	Tyr	Arg	Ile	His	Lys	Ser	Gly	Asp	Ile	Glu	Val
130						135				140				
Leu	Thr	Gly	Phe	Asn	Asn	Asn	Ala	Phe	Asp	Leu	Lys	Val	Pro	Lys
145					150					155				160
Leu	Leu	Asn	Pro	Ala	Gly	His	Ala	Ile	Tyr	Ile	Asp	Trp	Asn	Phe
				165					170					175
Ala	Thr	Gln	Pro	Arg	Leu	Asn	Arg	Ile	Tyr	Asp	Asp	Leu	Asp	Gly
			180					185					190	His
Asp	Ile	Pro	Leu	Leu	Asn	Leu	Glu	Tyr	Gln	Gly	Leu	Ile	Lys	Thr
		195					200					205		Ile
Leu	Thr	Leu	Phe	Pro	Gly	Gln	Lys	Glu	Gly	Tyr	Arg	Thr	Glu	Leu
		210				215					220			Arg
Phe	Leu	Asn	Arg	Gln	Leu	Asn	Ser	Ile	His	Asn	Phe	Ser	Leu	Gly
225					230					235				240
Glu	Asn	Pro	Leu	Thr	Trp	Ser	Phe	Gly	Tyr	Thr	Pro	Ile	Gly	Lys
				245					250					255
Gly	Ile	Leu	Gly	Gln	Trp	Ile	Thr	Ser	Met	Thr	Ala	Pro	Gly	Gly
			260					265					270	Leu
Lys	Glu	Thr	Val	Asn	Tyr	Ser	Asn	Asn	Asn	Gln	Gly	His	His	Phe
		275					280					285		Pro
Gln	Ser	Ala	Asn	Leu	Pro	Val	Leu	Pro	Tyr	Val	Thr	Leu	Met	Lys
		290				295					300			Gln
Val	Pro	Gly	Ala	Gly	Gln	Pro	Ala	Ile	Gln	Ala	Glu	Tyr	Ser	Tyr
305					310					315				320
Ser	His	Asn	Tyr	Val	Gly	Gly	Gly	Ser	Asn	Gly	Ile	Trp	Asn	Asn
				325					330					335
Leu	Asp	Asn	Leu	Tyr	Gly	Leu	Met	Thr	Glu	Tyr	Asn	Tyr	Gly	Ser
			340					345					350	Thr
Glu	Ser	Arg	Arg	Tyr	Lys	Asp	Lys	Glu	Gly	His	Asp	Gln	Ile	Val
		355					360					365		Arg
Ile	Glu	Arg	Thr	Tyr	Asn	Asn	Tyr	His	Leu	Leu	Thr	Ser	Glu	Cys
		370				375					380			Lys
Gln	Gln	Asn	Gly	Tyr	Ile	Gln	Thr	Thr	Glu	Thr	Ala	Tyr	Tyr	Ala
385					390					395				Ile
Ile	Gly	His	Asn	Phe	Asp	Ser	Gln	Pro	Ser	Gln	Phe	Gln	Leu	Pro
				405					410					415
Thr	Lys	Thr	Glu	Thr	Trp	Arg	Ser	Ala	Asp	Asn	Ser	Tyr	Arg	Ser
			420					425					430	Glu
Ile	Thr	Glu	Thr	Thr	Phe	Asp	Glu	Ser	Gly	Asn	Pro	Leu	Thr	Lys
		435					440					445		Val
Ile	Lys	Asp	Lys	Lys	Thr	Gln	Lys	Ile	Ile	Ser	Pro	Ser	Thr	His
		450				455					460			Trp
Glu	Tyr	Tyr	Pro	Pro	Ala	Gly	Glu	Val	Asp	Asn	Cys	Pro	Pro	Glu
465					470					475				Pro
Tyr	Gly	Phe	Thr	Arg	Phe	Val	Lys	Lys	Ile	Ile	Gln	Thr	Pro	Tyr
				485					490					495
Ser	Glu	Phe	Lys	Asp	Asp	Pro	Glu							

[illegible]

Thr Asp Leu Ala Thr Gly His Met Leu Thr Thr Thr Val Glu Phe Asp
 1045 1050 1055
 Gly Leu Asn Arg Glu Ile Gly Arg Lys Leu Cys Asp Ser Ser Gly His
 1060 1065 1070
 Thr Leu Asp Ile Gln Gln Ser Trp Leu Lys Thr Gln Gln Leu Ala Asn
 1075 1080 1085
 Arg Ile Val Lys Leu Asn Gly Val Leu Gln Arg Thr Glu Gln Tyr Ser
 1090 1095 1100
 Tyr Asp Ser Arg Asn Arg Leu Asn Gln Tyr Lys Cys Asp Gly Ala Glu
 1105 1110 1115 1120
 Cys Pro Thr Asp Lys Tyr Gly His Ser Ile Val Thr Gln Asn Phe Thr
 1125 1130 1135
 Tyr Asp Ile Tyr Gly Asn Ile Thr Ala Cys His Thr Thr Phe Ala Asp
 1140 1145 1150
 Gly Thr Glu Asp His Ala Thr Phe Lys Phe Ala Asn Pro Thr Asp Pro
 1155 1160 1165
 Cys Gln Leu Thr Glu Val His His Thr His Pro Asp Met Pro Asp Asn
 1170 1175 1180
 Ile Arg Leu Lys Tyr Asp Lys Ala Gly Arg Val Ile Asn Ile Thr Asp
 1185 1190 1195 1200
 Asn His Gly Asn Thr Glu Asn Phe Thr Tyr Asp Thr Leu Gly Arg Leu
 1205 1210 1215
 Gln Asn Gly Gln Gly Ser Val Tyr Gly Tyr Asp Pro Leu Asn Arg Leu
 1220 1225 1230
 Val Ser Gln Lys Thr Asp Thr Leu Asp Cys Glu Leu Tyr Tyr Arg Glu
 1235 1240 1245
 Thr Met Leu Val Asn Glu Val Arg Asn Gly Glu Met Ile Arg Leu Leu
 1250 1255 1260
 Arg Thr Gly Glu Thr Ile Ile Ala Gln Gln Arg Ala Ser Lys Val Leu
 1265 1270 1275 1280
 Leu Thr Gly Thr Asp Ser Gln Gln Ser Val Ile Leu Thr Ser Asp Lys
 1285 1290 1295
 Gln Asn Leu Ser Gln Glu Ala Tyr Ser Ala Tyr Gly Lys His Lys Ser
 1300 1305 1310
 Thr Ala Asn Asp Ala Ser Ile Leu Gly Tyr Asn Gly Glu Arg Ala Asp
 1315 1320 1325
 Pro Val Ser Gly Val Thr His Leu Gly Asn Gly Tyr Arg Ser Tyr Asp
 1330 1335 1340
 Pro Thr Leu Met Arg Phe His Thr Pro Asp Ser Leu Ser Pro Phe Gly
 1345 1350 1355 1360
 Ala Gly Gly Ile Asn Pro Tyr Ser Tyr Cys Leu Gly Asp Pro Ile Asn
 1365 1370 1375
 Arg Ser Asp Pro Ser Gly His Leu Ser Trp Gln Ala Trp Thr Gly Ile
 1380 1385 1390
 Gly Met Gly Ile Ala Gly Leu Leu Leu Thr Ile Ala Thr Gly Gly Met
 1395 1400 1405
 Ala Ile Ala Ala Ala Gly Gly Ile Ala Ala Ala Ile Ala Ser Thr Ser
 1410 1415 1420
 Thr Thr Ala Leu Ala Phe Gly Ala Leu Ser Val Thr Ser Asp Ile Thr
 1425 1430 1435 1440
 Ser Ile Val Ser Gly Ala Leu Glu Asp Ala Ser Pro Lys Ala Ser Ser
 1445 1450 1455
 Ile Leu Gly Trp Val Ser Met Gly Met Gly Ala Ala Gly Leu Ala Glu
 1460 1465 1470
 Ser Ala Ile Lys Gly Gly Thr Lys Leu Ala Thr His Leu Gly Ala Phe
 1475 1480 1485
 Ala Glu Asp Gly Glu Asn Ala Leu Leu Lys Ser Thr Ser Glu Ser Ser

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<210> 24
<211> 105
<212> PRT
<213> Xenorhabdus bovienii
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<210> 25
<211> 129
<212> PRT
<213> Xenorhabdus bovienii
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<400> 25
Ser Ala Gln Cys Ile Val Gly Lys Val Phe Arg Ile Ser Met Val Ile
 1          5          10          15
Ser Asp Ile Tyr Tyr Ser Thr Ser Leu Ile Ile Phe Gln Pro Asp Ile
          20          25          30
Ile Arg His Ile Trp Met Ser Val Tyr Leu Cys Gln Leu Ala Trp
      35          40          45

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Val Ser Trp Val Gly Lys Phe Glu Gly Ser Met Val Phe Cys Pro Ile
 50 55 60
 Cys Glu Cys Gly Val Thr Gly Gly Asp Ile Ala Ile Asp Ile Ile Ser
 65 70 75 80
 Lys Ile Leu Cys Asp Tyr Ala Met Ala Ile Phe Val Cys Arg Ala Phe
 85 90 95
 Arg Thr Val Thr Phe Ile Leu Val Gln Pro Ile Thr Gly Ile Val Arg
 100 105 110
 Val Leu Phe Cys Thr Leu Gln Tyr Ser Ile Gln Phe His Tyr Ser Ile
 115 120 125
 Cys

<210> 26
 <211> 141
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 26
 Pro Ser Ser Leu Arg Thr Ile Ser Leu Ser Lys Leu Leu Val Thr Pro
 1 5 10 15
 His Phe Ile Leu Glu Leu Ser Glu Val Asp Leu Ser Lys Ala Phe Ser
 20 25 30
 Pro Ser Ser Ala Asn Ala Pro Arg Cys Val Ala Ser Leu Val Pro Pro
 35 40 45
 Leu Met Ala Asp Ser Ala Asn Pro Ala Ala Pro Ile Pro Ile Glu Thr
 50 55 60
 His Pro Ser Ile Glu Asp Ala Phe Gly Glu Ala Ser Ser Ser Ala Pro
 65 70 75 80
 Leu Thr Ile Asp Val Ile Ser Asp Val Thr Leu Ser Ala Pro Asn Ala
 85 90 95
 Ser Ala Val Val Glu Val Glu Ala Ile Ala Ala Ala Ile Pro Pro Ala
 100 105 110
 Ala Ala Ile Ala Ile Pro Pro Val Ala Met Val Ser Ser Asn Pro Ala
 115 120 125
 Ile Pro Met Pro Ile Pro Val His Ala Cys Gln Leu Lys
 130 135 140

<210> 27
 <211> 101
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 27
 Ala His Cys His Ile Ala Leu Phe Pro Cys Trp His Asn Pro Gln Tyr
 1 5 10 15
 Cys Gln Gln His Pro Asp His His Ser Asn Cys His His Gln Phe Lys
 20 25 30
 Gln Glu Tyr Pro Pro Ser Arg Gln Arg Arg Glu Asn Ile Thr Leu Thr
 35 40 45
 Gln Leu Pro Ile Lys His Thr Gly Ile Glu Ala Gly Ser Gln Thr Asn
 50 55 60
 Arg Lys Arg Gln Thr Cys Met Phe Gln Arg Ala Asn Glu Ser Lys Val
 65 70 75 80
 His Gln Leu Gly Gln Asn Gln Gly Arg Asp Arg Asn Phe Tyr Trp Cys
 85 90 95
 Phe Asp Ile Leu Thr

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100

<210> 28
 <211> 117
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 28

Pro	Gln	Ser	Thr	Pro	Ser	Ser	Gln	Asn	Ser	Arg	Gln	Leu	Thr	Pro	Ala
1				5					10					15	
Glu	Ser	Ser	Gln	His	Gln	Lys	Gln	Lys	Ser	Asp	His	Ile	Glu	Ile	Met
			20					25					30		
Ile	Pro	Ser	Glu	Ala	Pro	Arg	Glu	Tyr	Arg	Glu	Gln	Leu	His	Lys	Ala
		35					40					45			
Thr	Pro	Ala	Arg	Asn	Arg	Asp	Val	Ala	Pro	Asn	Pro	Ser	Val	Phe	Asp
	50					55					60				
Ile	Leu	Arg	Asp	Tyr	His	Trp	Lys	Asn	Phe	Ser	Pro	Val	Lys	Ala	Ala
65					70					75					80
Lys	Ser	Ser	Leu	Thr	Pro	His	Pro	Val	His	Gln	Lys	Ala	Ile	Pro	Leu
				85					90					95	
Asn	Asp	Gln	Arg	Asn	Thr	Ser	Met	Lys	Gln	Ser	Leu	Lys	Pro	Glu	Met
			100					105						110	
Arg	Gln	Lys	Leu	Tyr											
			115												

<210> 29
 <211> 124
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 29

Gly	Lys	Asn	Cys	Ile	Asn	Asp	Gln	Gly	Asn	Leu	Pro	Asp	Arg	Tyr	Thr
1				5					10					15	
Gln	Asn	Cys	Arg	Pro	His	Leu	Thr	Asp	Asn	Pro	Pro	Tyr	Gly	Thr	Val
			20					25					30		
Thr	Glu	Arg	Asn	Pro	Arg	Gln	Tyr	Gln	His	Ala	Asp	Leu	Phe	Gln	Met
		35					40					45			
Arg	Lys	Leu	Ile	Gly	Gln	Leu	Gln	Asn	Pro	Ser	Gly	Asn	Asn	Gly	Pro
	50					55					60				
Thr	Gln	Arg	Gln	His	Trp	Arg	Ile	Ala	Ile	Arg	Ser	His	Lys	Gln	Cys
65					70					75					80
Lys	Asn	Asp	His	Thr	Asp	Ile	Glu	Gln	Cys	Arg	Ser	Lys	Ser	Arg	His
				85					90					95	
Arg	Lys	Ala	Val	Pro	Cys	Ile	Lys	Asn	Cys	Ala	Ser	Gln	Arg	Ser	Gln
			100					105						110	
Arg	Asn	Gln	Lys	Asp	Ile	Arg	Lys	Arg	Asn	Ser	Lys				
			115				120								

<210> 30
 <211> 515
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 30

Asn	Asn	Thr	Met	Asn	Leu	Leu	Lys	Ser	Leu	Ala	Ala	Val	Ser	Ser	Met
1				5					10					15	
Thr	Met	Phe	Ser	Arg	Val	Leu	Gly	Phe	Ile	Arg	Asp	Ala	Ile	Ile	Ala

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			20					25					30		
Arg	Ile	Phe	Gly	Ala	Gly	Met	Ala	Thr	Asp	Ala	Phe	Phe	Val	Ala	Phe
		35					40					45			
Lys	Leu	Pro	Asn	Leu	Leu	Arg	Arg	Ile	Phe	Ala	Glu	Gly	Ala	Phe	Ser
	50					55					60				
Gln	Ala	Phe	Val	Pro	Ile	Leu	Ala	Glu	Tyr	Lys	Asn	Gln	Gln	Gly	Asp
65					70					75					80
Glu	Ala	Thr	Arg	Thr	Phe	Ile	Ala	Tyr	Ile	Ser	Gly	Met	Leu	Thr	Leu
				85					90					95	
Ile	Leu	Ala	Ile	Val	Ser	Val	Ile	Gly	Val	Ile	Ala	Ala	Pro	Trp	Ile
				100				105					110		
Ile	Tyr	Val	Thr	Ala	Pro	Gly	Phe	Thr	Asp	Thr	Pro	Asp	Lys	Phe	Val
		115					120					125			
Leu	Thr	Arg	Asp	Leu	Leu	Arg	Ile	Thr	Phe	Pro	Tyr	Ile	Phe	Leu	Ile
	130					135					140				
Ser	Leu	Ala	Ser	Leu	Ala	Gly	Ala	Ile	Leu	Asn	Thr	Trp	Asn	Arg	Phe
145					150					155					160
Ser	Val	Pro	Ala	Phe	Ala	Pro	Thr	Leu	Leu	Asn	Val	Ser	Met	Ile	Ile
				165					170					175	
Phe	Ala	Leu	Phe	Val	Ala	Pro	Tyr	Cys	Asn	Pro	Pro	Val	Leu	Ala	Leu
			180					185					190		
Gly	Trp	Ala	Val	Val	Ala	Gly	Gly	Val	Leu	Gln	Leu	Ala	Tyr	Gln	Leu
		195					200					205			
Pro	His	Leu	Lys	Lys	Ile	Gly	Met	Leu	Val	Leu	Pro	Arg	Ile	Ser	Phe
	210					215					220				
Arg	Asp	Ser	Ala	Val	Trp	Arg	Val	Ile	Arg	Gln	Met	Gly	Pro	Ala	Ile
225				230						235					240
Leu	Gly	Val	Ser	Val	Gly	Gln	Ile	Ser	Leu	Ile	Ile	Asn	Thr	Ile	Phe
				245					250					255	
Ala	Ser	Phe	Leu	Val	Ser	Gly	Ser	Val	Ser	Trp	Met	Tyr	Tyr	Ala	Asp
			260					265					270		
Arg	Leu	Met	Glu	Leu	Pro	Ser	Gly	Val	Leu	Gly	Val	Ala	Leu	Gly	Thr
		275					280					285			
Ile	Leu	Leu	Pro	Ser	Leu	Ala	Lys	Ser	Phe	Ser	Ser	Gly	Asn	His	Glu
	290					295					300				
Glu	Tyr	Arg	Lys	Leu	Met	Asp	Trp	Gly	Leu	Arg	Leu	Cys	Phe	Leu	Leu
305					310					315					320
Ala	Leu	Pro	Cys	Ala	Val	Ala	Leu	Gly	Ile	Leu	Ala	Glu	Pro	Leu	Thr
				325					330					335	
Val	Ser	Leu	Phe	Gln	Tyr	Gly	His	Phe	Ser	Ala	Phe	Asp	Ala	Glu	Met
			340					345					350		
Thr	Gln	Arg	Ala	Leu	Ile	Ala	Tyr	Cys	Phe	Gly	Leu	Met	Gly	Leu	Ile
		355					360					365			
Val	Val	Lys	Val	Leu	Ala	Pro	Gly	Phe	Tyr	Ser	Arg	Gln	Asp	Ile	Lys
		370													

Leu Leu Arg Leu Met Gly Val Val Ile Ala Gly Ala Gly Ser Tyr Phe
 485 490 495
 Ala Val Leu Ala Leu Met Gly Phe Arg Leu Lys Asp Phe Ala His Arg
 500 505 510
 Gly Leu Gln
 515

<210> 31
 <211> 216
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 31
 Ala Ile Ile Leu Ile Arg Asp Lys Leu Ser Arg Ile Phe Ser Arg Gln
 1 5 10 15
 Ile Ser Gly Glu Gly Met Phe Gly Tyr Arg Ser Ala Ser Pro Lys Ile
 20 25 30
 Arg Phe Ile Thr Asp Arg Met Val Val Arg Leu Val Tyr Glu Arg Asp
 35 40 45
 Ala Tyr Arg Leu Ala Glu Tyr Tyr Ser Glu Asn Lys Asp Phe Leu Lys
 50 55 60
 Pro Trp Glu Pro Thr Arg Asp Gly Ser Phe Tyr Gln Pro Ser Gly Trp
 65 70 75 80
 Thr Asn Arg Leu Asn Tyr Ile Ala Glu Leu Gln Arg Gln Asn Ala Thr
 85 90 95
 Phe Asn Phe Val Leu Leu Asp Ser Asp Glu Arg Glu Ile Met Gly Val
 100 105 110
 Ala Asn Phe Thr Asn Val Val Arg Gly Ala Phe His Ser Cys Tyr Leu
 115 120 125
 Gly Tyr Ser Leu Ala Glu Lys Leu Gln Gly Gln Gly Leu Met Tyr Glu
 130 135 140
 Ala Leu Gln Pro Ala Ile Arg Tyr Met Gln Arg Tyr Gln Arg Met His
 145 150 155 160
 Arg Ile Met Ala Asn Tyr Met Pro His Asn His Arg Ser Gly Asn Leu
 165 170 175
 Leu Lys Lys Leu Gly Phe Glu Gln Glu Gly Tyr Ala Lys Asn Tyr Leu
 180 185 190
 Met Ile Asp Gly Val Trp Gln Asp His Val Leu Thr Ala Leu Thr Asp
 195 200 205
 Asp Ala Trp Gly Lys Val Gly Leu
 210 215

<210> 32
 <211> 404
 <212> PRT
 <213> Xenorhabdus bovienii

<400> 32
 Trp Cys Ala Met Ser Leu Val Ser Gln Ala Arg Ser Leu Gly Lys Tyr
 1 5 10 15
 Phe Leu Leu Phe Asp Asn Leu Leu Val Val Leu Gly Phe Phe Val Val
 20 25 30
 Phe Pro Leu Ile Ser Ile Arg Phe Val Glu Gln Leu Gly Trp Ala Ala
 35 40 45
 Leu Ile Val Gly Phe Ala Leu Gly Leu Arg Gln Leu Val Gln Gln Gly
 50 55 60
 Leu Gly Ile Phe Gly Gly Ala Ile Ala Asp Arg Phe Gly Ala Lys Pro

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65 70 75 80
 Met Ile Val Thr Gly Met Leu Leu Arg Ala Leu Gly Phe Ala Leu Met
 85 90 95
 Ala Met Ala His Glu Pro Trp Ile Leu Leu Ser Cys Val Leu Ser
 100 105 110
 Gly Leu Gly Gly Thr Leu Phe Asp Pro Pro Arg Ala Ala Leu Val Ile
 115 120 125
 Lys Leu Thr Arg Pro His Glu Arg Gly Arg Phe Tyr Ser Ile Leu Met
 130 135 140
 Met Gln Asp Ser Ala Gly Ala Val Val Gly Ala Leu Ile Gly Ser Trp
 145 150 155 160
 Leu Leu Gln Tyr Asp Phe Asn Ile Val Cys Trp Ile Gly Ala Ser Ile
 165 170 175
 Phe Val Leu Ala Ala Leu Phe Asn Ala Trp Leu Leu Pro Ala Tyr Arg
 180 185 190
 Ile Ser Thr Ile Arg Thr Pro Ile Lys Glu Gly Met Met Arg Val Ile
 195 200 205
 Arg Asp Arg Arg Phe Leu Tyr Tyr Val Leu Thr Leu Thr Gly Tyr Phe
 210 215 220
 Val Leu Ser Val Gln Val Met Leu Met Phe Pro Ile Ile Ile His Glu
 225 230 235 240
 Ile Thr Gly Thr Pro Thr Ala Val Lys Trp Met Tyr Ala Ile Glu Thr
 245 250 255
 Ala Ile Ser Leu Thr Leu Leu Tyr Pro Ile Ala Arg Trp Ser Glu Lys
 260 265 270
 His Phe Arg Leu Glu Gln Arg Leu Met Ala Gly Leu Phe Leu Met Ser
 275 280 285
 Ile Cys Met Phe Pro Ile Gly Trp Val Asn Gln Leu His Thr Leu Phe
 290 295 300
 Gly Leu Leu Cys Leu Phe Tyr Leu Gly Leu Val Thr Ala Asp Pro Ala
 305 310 315 320
 Arg Glu Thr Leu Ser Ala Ser Leu Ser Asp Pro Arg Ala Arg Gly Ser
 325 330 335
 Tyr Met Gly Phe Ser Arg Leu Gly Leu Ala Leu Gly Gly Ala Ile Gly
 340 345 350
 Tyr Thr Gly Gly Gly Trp Leu Tyr Asp Thr Gly Arg Asp Leu Asn Met
 355 360 365
 Pro Gln Leu Pro Trp Ile Leu Leu Gly Leu Ser Gly Leu Ile Thr Ile
 370 375 380
 Tyr Ala Leu His Arg Gln Phe Asn Gln Lys Lys Ile Asp Pro Val Met
 385 390 395 400
 Leu Gly Arg His

<210> 33

<211> 191

<212> PRT

<213> Xenorhabdus bovienii

<400> 33

Lys Gly Ala Asn Met Lys Arg Phe Phe Leu Gly Ala Ala Leu Val Leu
 1 5 10 15
 Val Gly Leu Val Ser Gly Cys Asp Gln Phe Lys Asp Phe Ser Ile Asn
 20 25 30
 Glu Gly Leu Met Asn Asp Tyr Leu Lys Lys Val His Tyr Gln Lys
 35 40 45
 Lys Ile Ser Ile Pro Gly Ile Ala Asn Ala Asn Ile Thr Leu Gly Asp

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<210> 34
<211> 205
<212> PRT
<213> Xenorhabdus bovienii
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<210> 35
<211> 315
<212> PRT
<213> Xenorhabdus bovienii
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<210> 36
<211> 132
<212> PRT
<213> Xenorhabdus bovienii
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<400> 36																
Lys	Thr	Ser	Gln	Gly	Phe	Thr	Ser	Thr	Thr	Cys	Ser	Asn	Gly	Asn	Val	
1				5					10					15		
Leu	Lys	Ile	Cys	Gly	Leu	Ile	Thr	Pro	Cys	Ser	Ser	Leu	Ile	Gln	Arg	
			20					25					30			
Thr	Tyr	Pro	Asn	Asn	Met	Thr	Ile	Gly	Ile	Phe	Ser	Lys	Glu	Ser	Thr	
		35					40					45				
Ala	Lys	Asn	Phe	Gly	Met	Gly	Phe	Leu	Tyr	Tyr	Phe	Asp	Leu	Arg	Val	
	50					55					60					
Leu	Ser	Pro	Phe	Phe	Lys	Ala	Pro	Ile	Asn	Ile	Phe	Thr	Gly	Trp	Gln	
65					70					75					80	

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<210> 37
<211> 289
<212> PRT
<213> Xenorhabdus bovienii
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<210>	38
<211>	270
<212>	PRT

<213> Xenorhabdus bovienii

<400> 38

Lys Gly Asn Gln Ile Thr Met Ile Leu Tyr Lys Gly Ser Lys Asn Tyr
 1 5 10 15
 Leu Phe Asn Gln Leu Asn Tyr Asp Ser Cys Val Leu Leu Glu Val Asp
 20 25 30
 Glu Ser Val Asn Leu Asn Gly Trp Asp Glu Leu Ser Arg Ala Gln Arg
 35 40 45
 Leu Leu Phe Leu Met Glu Ile Leu Arg Arg Tyr His Phe Pro Val Gln
 50 55 60
 Gly Lys Val Leu Ala Gln Lys Leu Asn Ile Ser Leu Arg Thr Leu Tyr
 65 70 75 80
 Arg Asp Ile Ala Ser Leu Gln Ala Gln Gly Ala Ile Ile Glu Gly Glu
 85 90 95
 Pro Gly Ile Gly Tyr Val Leu Arg Pro Gly Phe Val Leu Pro Pro Leu
 100 105 110
 Met Phe Thr Gln Asn Glu Ile Glu Ala Leu Ala Leu Gly Ala Asn Trp
 115 120 125
 Val Ala Lys Arg Ala Asp Pro Gln Leu Lys Glu Ser Ala Asn Asn Ala
 130 135 140
 Ile Ser Lys Ile Ala Ala Val Ile Pro Ala Glu Leu Lys Gln Met Leu
 145 150 155 160
 Glu Ala Ser Ser Leu Leu Ile Gly Pro Ala Ala Thr Ala Val Gln Pro
 165 170 175
 Val Val Glu Ile Gln Gln Ile Arg Gln Ala Ile Asn Thr Arg His Lys
 180 185 190
 Ile Thr Leu Ala Tyr Leu Asp Ile Lys Asp Ile Pro Ser Glu Arg Thr
 195 200 205
 Ile Trp Pro Phe Ala Leu Gly Tyr Phe Glu Asn Ile Ser Ile Val Ile
 210 215 220
 Gly Trp Cys Glu Leu Arg Glu Glu Phe Arg His Phe Arg Ser Asp Arg
 225 230 235 240
 Ile Met Arg Leu Lys Ile Glu Asn Gln Cys Tyr Pro Arg Ser Arg Gln
 245 250 255
 Val Leu Leu Lys Glu Trp Arg Ala Met Glu Lys Ile Ser Arg
 260 265 270

<210> 39

<211> 209

<212> PRT

<213> Xenorhabdus bovienii

<400> 39

Arg Lys Met Thr Ile Tyr Asp Leu Lys Pro Arg Phe Gln Asn Leu Leu
 1 5 10 15
 Arg Pro Ile Val Ile Tyr Leu Tyr Lys Gln Gly Ile Thr Ala Asn Gln
 20 25 30
 Val Thr Leu Thr Ala Leu Phe Leu Ser Ile Phe Ala Gly Ser Leu Leu
 35 40 45
 Ser Leu Phe Pro Ser Pro His Leu Tyr Trp Leu Leu Pro Val Phe Leu
 50 55 60
 Phe Ile Arg Met Ala Leu Asn Ala Ile Asp Gly Met Leu Ala Arg Glu
 65 70 75 80
 His Asn Gln Lys Ser His Leu Gly Ala Ile Tyr Asn Glu Leu Gly Asp
 85 90 95
 Val Ile Ser Asp Val Ala Leu Tyr Leu Pro Phe Cys Leu Leu Pro Asp

100 105 110
 Val Asn Ser Leu Ser Leu Leu Ile Ile Leu Phe Leu Thr Ile Leu Thr
 115 120 125
 Glu Phe Ile Gly Val Leu Ala Gln Thr Ile Gly Ala Ser Arg Arg Tyr
 130 135 140
 Asp Gly Pro Ile Gly Lys Ser Asp Arg Ala Phe Ile Phe Gly Ala Tyr
 145 150 155 160
 Gly Leu Ile Ile Ala Ile Phe Pro Leu Ala Leu Gly Trp Ser Ile Ser
 165 170 175
 Leu Phe Ala Phe Met Ile Ile Leu Leu Leu Val Thr Cys Tyr Gln Arg
 180 185 190
 Val Val Lys Ala Leu Arg Glu Ile Arg Leu Ala Glu Gln Ser His Ser
 195 200 205

Lys

<210> 40

<211> 592

<212> PRT

<213> Xenorhabdus bovienii

<400> 40

Gly Val Asn Met Thr Pro Gln Leu Asp Gln Arg Ile Ala Glu Glu His
 1 5 10 15
 Tyr Phe Thr Thr Ser Asp Asn Ala Ser Leu Phe Tyr Arg Tyr Trp Pro
 20 25 30
 Gln Gln Gln Ala Asn Pro Asp Arg Ala Ile Ile Ile Phe His Arg Gly
 35 40 45
 His Glu His Ser Gly Arg Ile Gln His Val Val Asp Gly Leu Asp Leu
 50 55 60
 Pro Asp Val Pro Met Phe Ala Trp Asp Ala Arg Gly His Gly Lys Thr
 65 70 75 80
 Glu Gly Pro Arg Gly Tyr Ser Pro Ser Met Gly Thr Ser Ile Arg Asp
 85 90 95
 Val Asp Glu Phe Val Arg Phe Ile Ala Thr Gln Tyr Gly Ile Ala Met
 100 105 110
 Glu Asn Ile Val Val Ile Gly Gln Ser Val Gly Ala Val Leu Val Ser
 115 120 125
 Ala Trp Val His Asp Tyr Ala Pro Lys Ile Arg Ala Met Ile Leu Ala
 130 135 140
 Ala Pro Ala Phe Asp Ile Lys Leu Tyr Ile Pro Phe Ala Thr Gln Gly
 145 150 155 160
 Leu Gln Leu Met Gln Lys Ala Arg Gly Ile Phe Phe Val Asn Ser Tyr
 165 170 175
 Val Lys Ala Arg Tyr Leu Thr His Asp Glu Thr Arg Ile Ala Ser Tyr
 180 185 190
 Asn Ser Asp Pro Leu Ile Thr Arg Glu Ile Ala Val Asn Ile Leu Leu
 195 200 205
 Asp Leu Tyr Gln Thr Ala Glu Arg Val Val Lys Asp Ala Ala Ala Ile
 210 215 220
 Thr Leu Pro Thr Leu Leu Phe Ile Ser Gly Ser Asp Tyr Val Val Asn
 225 230 235 240
 Lys Lys Pro Gln His Gln Phe Tyr Gln Gln Leu Asn Thr Pro Ile Lys
 245 250 255
 Glu Lys His Val Met Asp Gly Phe Tyr His Asp Thr Leu Gly Glu Lys
 260 265 270
 Asp Arg His Leu Val Phe Asp Lys Ile Arg Val Phe Ile Glu Arg Ile

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<210> 41
<211> 121
<212> PRT
<213> Xenorhabdus bovienii
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<400>	41														
His	His	Asn	Ser	Ile	Asn	Val	Leu	Leu	Lys	Asn	Ile	Ile	Ser	Pro	His
1				5					10					15	
Gln	Ile	Met	Leu	Leu	Cys	Phe	Thr	Val	Thr	Gly	His	Asn	Asn	Arg	Pro
			20					25					30		
Ile	Gln	Thr	Glu	Arg	Ser	Leu	Phe	Phe	Thr	Val	Val	Met	Ser	Thr	Gln
		35					40					45			
Asp	Val	Ser	Ser	Met	Ser	Leu	Thr	Asp	Ser	Ile	Cys	Leu	Met	Phe	Leu
	50					55					60				
Cys	Ser	Arg	Gly	Met	Pro	Val	Asp	Thr	Val	Arg	Gln	Lys	Gly	Arg	Ala
65					70					75					80
Val	Thr	Ala	His	Pro	Trp	Glu	Arg	Arg	Phe	Val	Met	Leu	Met	Asn	Leu


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<210> 42
<211> 444
<212> PRT
<213> Xenorhabdus bovienii
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<400>	42																
Ile	Asn	Lys	Tyr	Lys	Met	Glu	His	His	Met	His	Ser	Ser	Leu	Asp	Ser		
1				5					10					15			
Arg	Arg	Arg	Leu	Trp	Leu	Thr	Gly	Val	Ile	Trp	Leu	Leu	Phe	Leu	Ala		
			20					25					30				
Pro	Phe	Phe	Phe	Leu	Thr	Tyr	Gly	Gln	Val	Asn	Gln	Phe	Thr	Ala	Gln		
		35					40					45					
Arg	Ser	Asp	Val	Gly	Thr	Val	Met	Phe	Gly	Trp	Glu	His	Asn	Ile	Pro		
	50					55					60						
Phe	Trp	Ser	Trp	Ser	Ile	Ile	Pro	Tyr	Trp	Ser	Ile	Asp	Leu	Phe	Tyr		
65					70					75					80		
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Leu	Phe	Pro	Leu	Lys	Phe	Ser	Phe	Ser	Arg	Pro	Thr	Thr	Glu	Gly	Leu		
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Phe	Gly	Trp	Leu	Phe	Asn	Gln	Leu	Glu	Leu	Phe	Asp	Leu	Pro	Tyr	Asn		
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Gln	Ala	Pro	Ser	Leu	His	Ile	Ile	Leu	Leu	Trp	Leu	Leu	Trp	Leu	Arg		
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Tyr	Ser	Ala	Tyr	Val	Ser	Gly	Tyr	Trp	Arg	Gly	Leu	Leu	His	Ile	Trp		
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Ser	Val	Leu	Ile	Ala	Leu	Ser	Val	Leu	Thr	Thr	Trp	Gln	His	His	Phe		
			180					185					190				
Ile	Asp	Val	Leu	Thr	Gly	Phe	Ala	Val	Gly	Val	Ile	Leu	Ser	Tyr	Leu		
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Leu	Pro	Val	Ser	Tyr	Arg	Trp	Arg	Trp	Gln	Pro	Asn	Gln	Asp	Arg	Tyr		
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Ala	Arg	Lys	Leu	Phe	Gly	Tyr	Tyr	Leu	Thr	Gly	Ser	Ala	Leu	Phe	Ala		
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				245					250					255			
Ala	Val	Ser	Leu	Leu	Met	Ile	Ala	Leu	Gly	Tyr	Ala	Gly	Leu	Gly	Ser		
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Trp	Leu	Leu	Ala	Pro	Tyr	Gln	Leu	Gly	Ala	Trp	Leu	Ser	Tyr	Leu	Trp		
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<213> Xenorhabdus bovienii
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 20 25 30
 Ala Glu Arg Ala Ile Ala Ala Val Met Pro Pro Lys Lys Gly
 35 40 45
 Ala Ala Ile Leu Pro Asn Pro Trp Pro Ser Ser Ser Pro Leu Glu Trp
 50 55 60
 Cys Phe Phe Pro Val Ile Pro Ser Arg Ile Thr Ala His Ser Asn Asp
 65 70 75 80
 Ser Ile Ala Pro Ser Met Ala Ile Glu Asn Ala Ala Gly Ser Asn Ala
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 Asp Thr Val Phe Gln Leu Ile Ser Arg Glu Cys Val Ser Gly Lys Phe
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 His Gly Arg Thr Asn Trp Gly Arg Met Gly Gly Met Pro
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<210> 46

<211> 161

<212> PRT

<213> Xenorhabdus bovienii

<400> 46

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 Thr Ser Ser Leu Thr Ser Ala Glu Lys Gly Leu Leu Val Val Arg Ile
 35 40 45
 Asn Gly Pro Leu Phe Phe Ala Ala Ala Glu Arg Ile Phe Ala Glu Leu
 50 55 60
 Arg Glu Lys Ser Ala Asp Tyr Gln Thr Ile Ile Met Gln Trp Asp Ala
 65 70 75 80
 Val Pro Val Leu Asp Ala Gly Gly Leu His Ala Phe Gln Gly Phe Val
 85 90 95
 Arg Glu Leu Gly Lys Glu Lys His Ile Val Val Cys Asp Ile Pro Phe
 100 105 110
 Gln Pro Leu Lys Thr Leu Ala Arg Ala Lys Val Met Pro Ile Glu Gly
 115 120 125
 Glu Leu Ser Phe Tyr Ala Thr Leu Pro Lys Ala Leu Lys Glu Met Ala
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 Val Asp Tyr Thr Pro Glu Val Cys Ala Ser Ser Glu Lys Ile Gln Gly
 145 150 155 160
 Gln

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<211> 173

<212> PRT

<213> Xenorhabdus bovienii

<400> 47

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09889974-10304

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<213> Xenorhabdus bovienii
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Gly	Asp	Ile 35	Lys	Val	Ala	Asn 40	Asp	Leu	Pro	Phe	Val 45	Leu	Phe	Gly	Gly	
Met	Asn 50	Val	Leu	Glu	Ser	Arg 55	Asp	Leu	Ala	Met 60	Arg	Ile	Cys	Glu	His	
Tyr 65	Val	Thr	Val	Thr 70	Gln	Lys	Leu	Gly	Ile 75	Pro	Tyr	Val	Phe	Lys	Ala 80	
Ser	Phe	Asp	Lys 85	Ala	Asn	Arg	Ser	Ser 90	Ile	Arg	Ser	Tyr	Arg 95	Gly	Pro	
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Gly	Val	Lys 115	Ile	Ile	Thr	Asp 120	Val	His	Glu	Pro	Ala 125	Gln	Ala	Gln	Pro	
Val	Ala 130	Asp	Val	Val	Asp 135	Val	Ile	Gln	Leu	Pro 140	Ala	Phe	Leu	Ala	Arg	
Gln 145	Thr	Asp	Leu	Val 150	Glu	Ala	Met	Ala	Lys	Thr 155	Gly	Ala	Val	Ile	Asn 160	
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Glu	Lys	Phe	Lys 180	Glu	Gly	Gly	Asn 185	Asp	Gln	Val	Ile	Leu	Cys 190	Asp	Arg	
Gly	Ser 195	Asn	Phe	Gly	Tyr	Asp 200	Asn	Leu	Val	Val 205	Asp	Met	Leu	Gly	Phe	
Gly 210	Val	Met	Gln	Gln	Ala 215	Thr	Gln	Gly	Ala	Pro 220	Val	Ile	Phe	Asp	Val	
Thr 225	His	Ala	Leu	Gln 230	Cys	Arg	Asp	Pro	Leu	Gly 235	Ala	Ala	Ser	Gly	Gly 240	
Arg	Arg	Ala	Gln 245	Val	Ala	Glu	Leu	Ala 250	Arg	Ala	Gly	Met	Ala 255	Val	Gly	
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<213> Xenorhabdus bovienii
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 35 40 45
 Ser Pro Lys Arg Asp Ala Glu Ile Leu Leu Gly Tyr Val Thr Gly Arg
 50 55 60
 Ser Arg Thr Tyr Leu Ile Ala Phe Asp Glu Thr Leu Ile Ser Ser Glu
 65 70 75 80
 Glu Leu His Gln Leu Asp Ser Leu Leu Val Arg Arg Ile Gln Gly Glu
 85 90 95
 Pro Val Ala Tyr Ile Ile Gly Glu Arg Glu Phe Trp Ser Leu Pro Phe
 100 105 110
 Ala Val Ser Pro Ala Thr Leu Ile Pro Arg Pro Asp Thr Glu Cys Leu
 115 120 125
 Val Glu Lys Ala Leu Glu Leu Leu Pro Asp Ser Pro Ala Arg Ile Leu
 130 135 140
 Asp Leu Gly Thr Gly Thr Gly Ala Ile Ala Leu Ala Leu Ala Ser Glu
 145 150 155 160
 Arg Asn Asp Cys Tyr Val Thr Gly Val Asp Ile Asn Ser Asp Ala Val
 165 170 175
 Met Leu Ala Gln His Asn Ala Glu Lys Asn Ala Gly Lys Leu Ala Ile
 180 185 190
 His Asn Val Asn Phe Leu Gln Ser Glu Trp Phe Ala Ala Val Gly Asn
 195 200 205
 Gln Gln Phe Asp Met Ile Val Ser Asn Pro Pro Tyr Ile Asp Glu Arg
 210 215 220
 Asp Pro His Leu Gln Glu Gly Asp Ile Arg Phe Glu Pro Ala Thr Ala
 225 230 235 240
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 245 250 255
 Gln Ala Arg His Phe Leu Ser Pro Asn Gly Trp Leu Leu Leu Glu His
 260 265 270
 Gly Trp Lys Gln Gly Thr Val Val Arg Asn Leu Phe Leu Glu Lys Gly
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 Thr Ile Gly Arg Trp Asn Lys Asn Glu Thr His Ser
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<211> 289

<212> PRT

<213> Xenorhabdus bovienii

<400> 51

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 Asp Pro Asp Asp Glu Arg Asn Cys Phe Leu Glu Val Arg Ala Gly Thr
 35 40 45
 Gly Gly Asp Glu Ala Ala Ile Phe Ala Gly Asp Leu Phe Arg Met Tyr
 50 55 60
 Ser Arg Tyr Ala Glu Ala Arg Arg Trp Arg Val Glu Ile Ile Ser Ala
 65 70 75 80

T000001-42000000

Asn Glu Gly Glu His Gly Gly Tyr Lys Glu Val Ile Ala Lys Val Ser
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 Gly Asp Gln Val Tyr Gly His Leu Lys Phe Glu Ser Gly Gly His Arg
 100 105 110
 Val Gln Arg Val Pro Glu Thr Glu Ser Gln Gly Arg Ile His Thr Ser
 115 120 125
 Ala Cys Thr Val Ala Val Met Pro Glu Ile Pro Glu Ala Glu Leu Pro
 130 135 140
 Asp Ile Ser Pro Gly Asp Leu Lys Ile Asp Thr Phe Arg Ser Ser Gly
 145 150 155 160
 Ala Gly Gly Gln His Val Asn Thr Thr Asp Ser Ala Ile Arg Ile Thr
 165 170 175
 His Leu Pro Thr Gly Ile Val Val Glu Cys Gln Asp Glu Arg Ser Gln
 180 185 190
 His Lys Asn Lys Ala Lys Ala Met Ser Val Leu Ala Ala Arg Ile Arg
 195 200 205
 Ala Ala Glu Met Arg Lys Arg Gln Glu Val Glu Ala Ser Glu Arg Arg
 210 215 220
 Asn Leu Leu Gly Ser Gly Asp Arg Ser Asp Arg Asn Arg Thr Tyr Asn
 225 230 235 240
 Phe Pro Gln Gly Arg Val Thr Asp His Arg Ile Asn Leu Thr Leu Tyr
 245 250 255
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<212> DNA

<213> Xenorhabdus bovienii

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